



Opportunities for Facility-Enabled Science at the DOE Joint Genome Institute (JGI)

Yasuo Yoshikuni, Ph.D.
Head, DNA Synthesis Science Program
12/20/2024



Overview of Research Focus Areas



DNA Synthesis Science Program

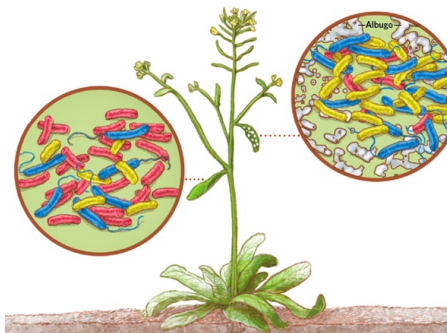


Yoshikuni Research Group

Biomass-side

Product-side

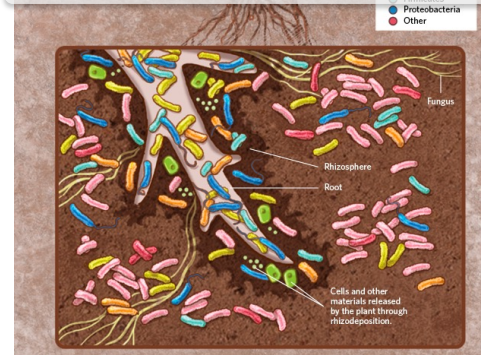
1. Plant N-utilization



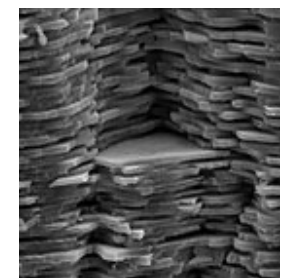
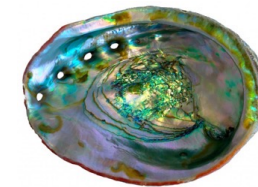
3. Biofuel and Bioproducts



2. Microbiome engineering



4. Hybrid Biomaterials



Overview of Research Focus Areas

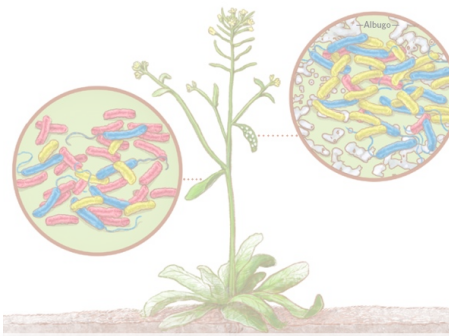
DNA Synthesis Science Program



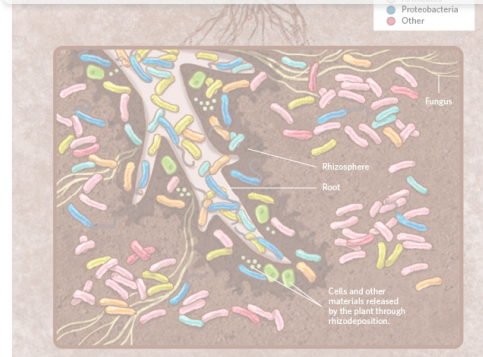
Yoshikuni Research Group

Biomass-side

1. Plant N-utilization

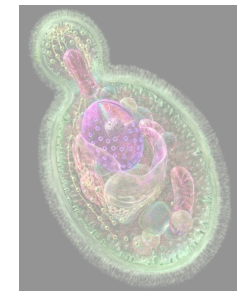


2. Microbiome engineering

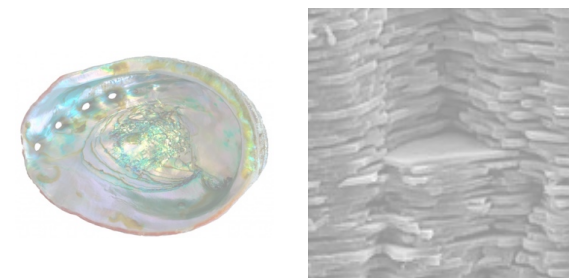


Product-side

3. Biofuel and Bioproducts



4. Hybrid Biomaterials



The DOE Joint Genome Institute



Integrative Genomics Building (IGB)



JGI MISSION:

To provide the global research community with **free access** to the most advanced integrative genome science capabilities in support of the DOE energy & environmental research mission



U.S. Department of Energy Office of Science User Facility

- JGI established in 1997, User facility from 2004
- Located at Lawrence Berkeley National Laboratory
- ~285 staff; \$91.6 M annual funding
- 2,038 Global Primary Users; >10,000 Data Users



DNA Synthesis Science Program



Mission Statement

To harness the power of DNA synthesis, strain engineering, and biosystems design for DOE mission relevant discovery and applications

The screenshot shows the JGI website's 'User Programs' section. The main heading is 'CSP Functional Genomics'. Below it, there is a 'SUBMIT' button and a section titled 'About This Call'. The text describes the call's purpose: to enable users to perform state-of-the-art functional genomics research and to help them translate genomic information into biological function. It mentions that proposals should be submitted to the JGI at any time using a simple web-based form. The text also lists several key personnel: Yasuo Yoshikuni (DNA synthesis program head), Trent Northen (metabolomics group lead), Tanja Woyke (Interim Deputy for User Programs), and Miranda Harmon-Smith (project manager). A blue arrow points from this text to the 'Key achievements' section. At the bottom of the screenshot, it says 'Current Call (OPEN) 1/30/2025 (DEADLINE)'.

Large-scale synbio projects

- 500 kbp DNA synthesis, construct assembly
- Fast-track metabolic engineering
- sgRNA library
- Strain engineering

etc....

Key achievements

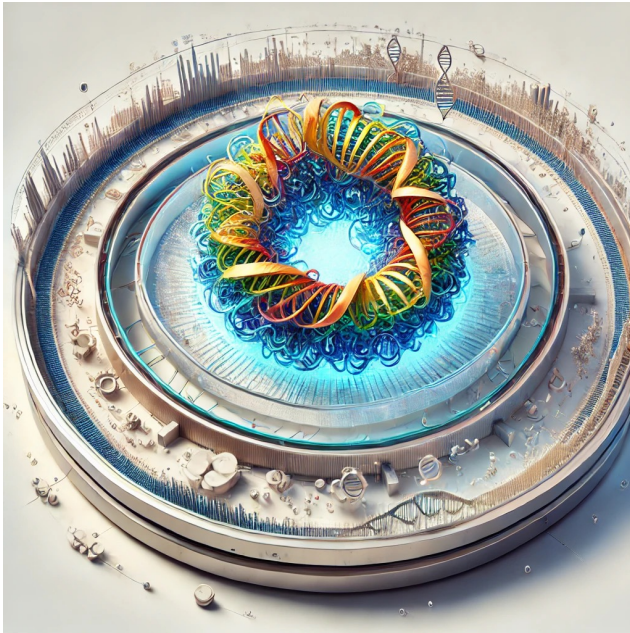
- > 300 projects
 - > 200 publications
- Including Nature, Science, Nature Sister Journals*



1/30/2025 (DEADLINE)

Strategic Focus Areas – Implementation of 2018 Plan

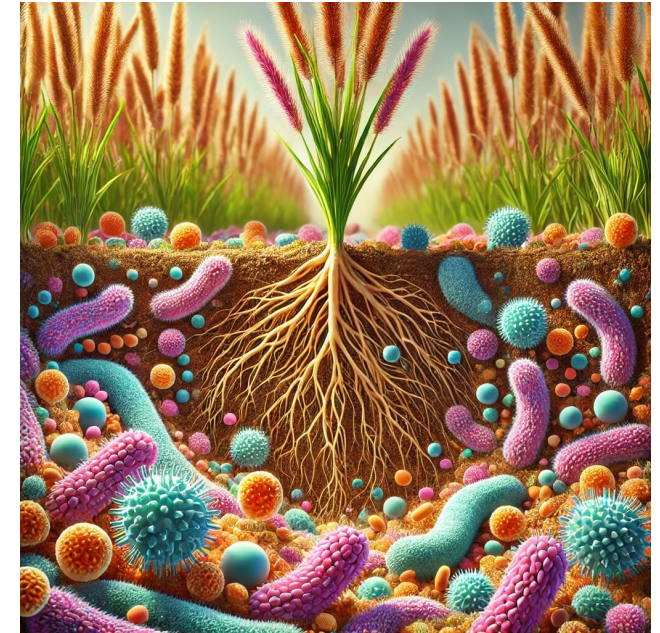
1. Genomes to structure and function



2. High-throughput functional genomics



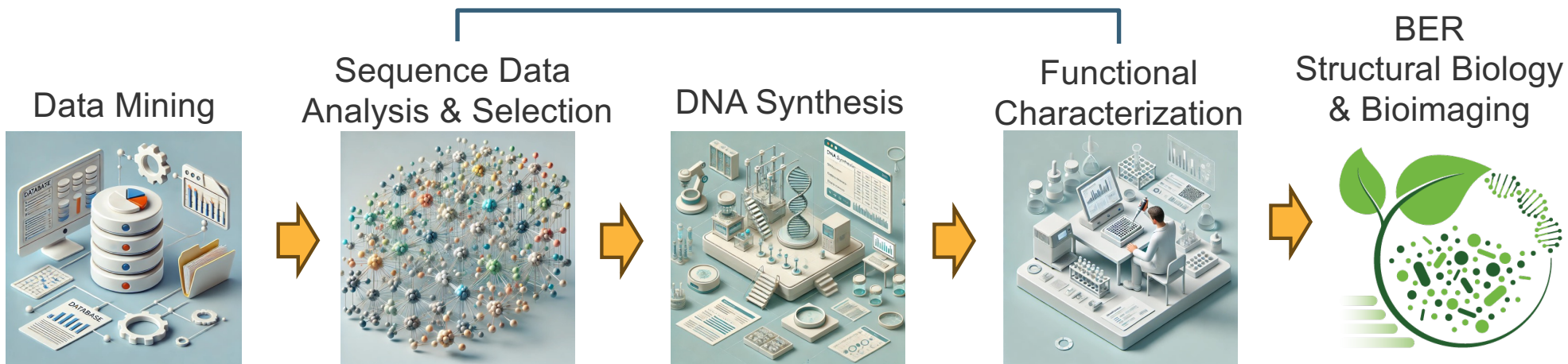
3. Microbe-microbe and plant-microbe interactions



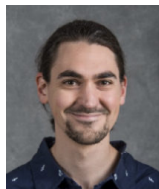
Genomes to Structure and Function

The program oversees scientific communications with Users and facilitate capability integration

User-Driven



Natalia Ivanova



Simon Roux

Platform



Ian Blaby

Metabolomics

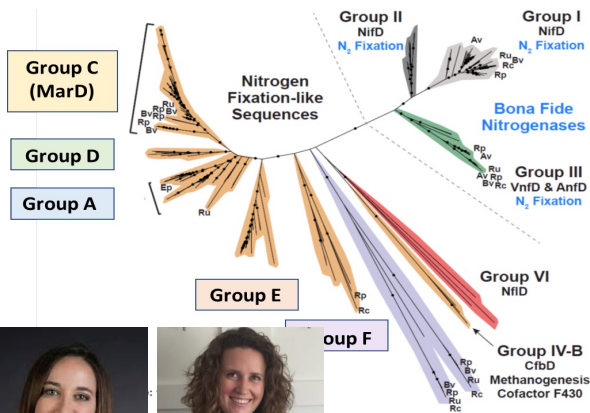


Trent Northen



Katherine Louie



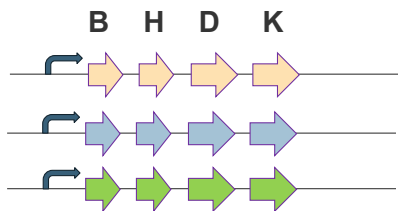


Kelly Wrighton



Emiley Eloie-Fadrosch

Gene Cluster Assembly



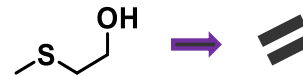
Ian Blaby

Ethylene Group

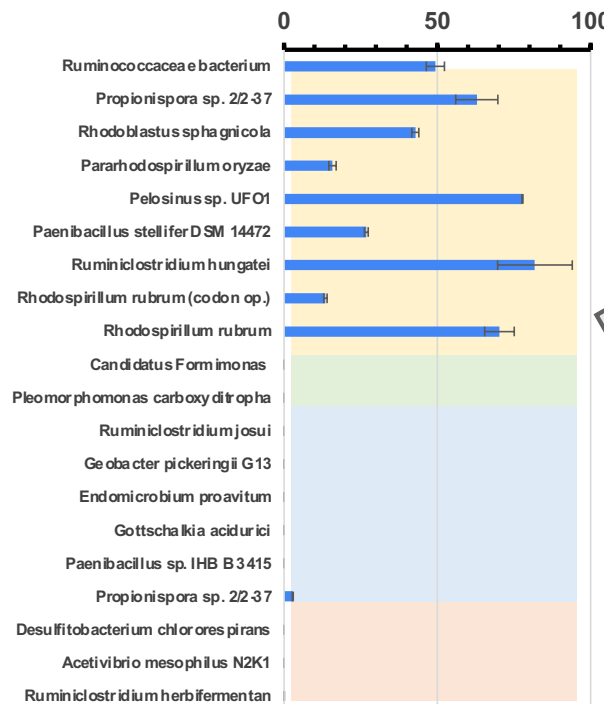
Screen for functional Ethylene Enzymes



Justin North



μmol/L/O.D.



MarD
MarK

Structure Determination



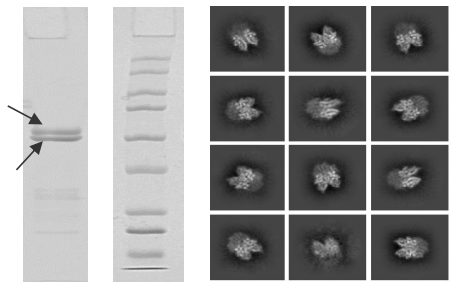
Brookhaven National Laboratory
Sean McSweeney



Brookhaven National Laboratory
Dale Kreidler

Sean McSweeney

Dale Kreidler



Cryo EM



Under review

Science Portfolio 1

Other Examples

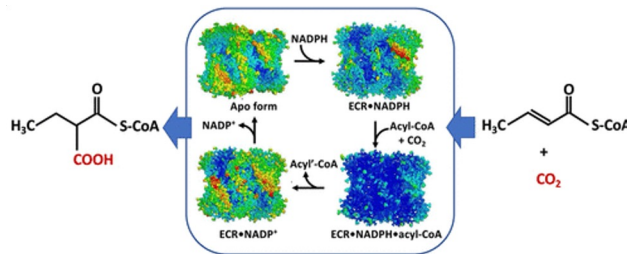
Challenge: Integration of capabilities across facilities

Results: Demonstrated the power of capability integration

ACS central science

Intersubunit Coupling Enables Fast CO₂-Fixation by Reductive Carboxylases

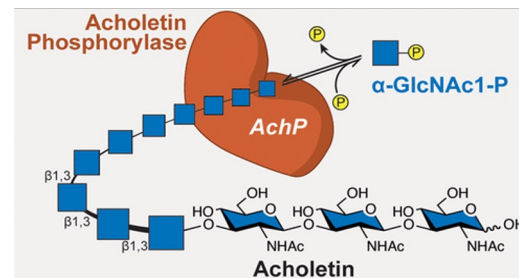
Hasan DeMirci^{1,2}, Yashas Rao¹, Gabriele M. Stoffel¹, Bastian Vögeli¹, Kristina Schell¹, Aharon Gomez¹, Alexander Batyuk¹, Cornelius Gati¹, Raymond G. Sierra¹, Mark S. Hunter¹, E. Han Dao¹, Hali I. Ciftci¹, Brandon Hayes¹, Fredric Poitevin¹, Po-Nan Li¹, Manat Kaur¹, Kensuke Tono¹, David Adrian Saez¹, Samuel Deutsch¹, Yasuo Yoshikuni¹, Helmut Grubmüller¹, Tobias J. Erb^{1*}, Esteban Vöhringer-Martinez^{1,2} and Soichi Wakatsuki^{1*}



ACS central science

A Synthetic Gene Library Yields a Previously Unknown Glycoside Phosphorylase That Degrades and Assembles Poly-β-1,3-GlcNAc, Completing the Suite of β-Linked GlcNAc Polysaccharides

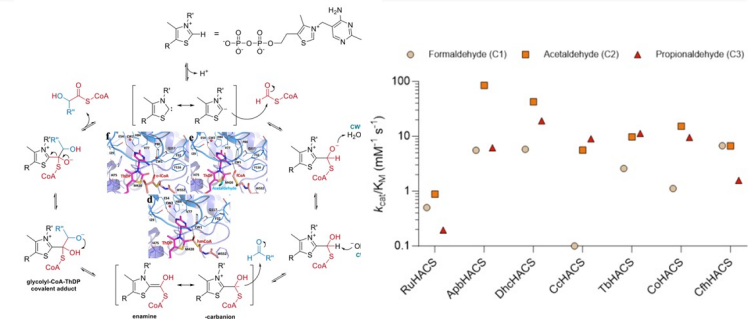
Spencer S. Macdonald¹, Jose H. Pereira¹, Feng Liu¹, Gregor Tegli¹, Andy DeGiovanni¹, Jacob F. Wardman¹, Samuel Deutsch¹, Yasuo Yoshikuni¹, Paul D. Adams¹, and Stephen G. Withers^{1*}



communications biology

Revealing reaction intermediates in one-carbon elongation by thiamine diphosphate/CoA-dependent enzyme family

Yongchang Kim^{1,2}, Seung Heon Lee¹, Priyanka Gadi¹, Maren Nattermann¹, Natalia Matveeva¹, Michael Endler¹, Jing Chen¹, Philipp Weidmann¹, Yang He¹, Daniel G. Marshall¹, Yasuo Yoshikuni^{1,2}, Tobias J. Erb^{1,2}, Ramon Gonzalez^{1,2,3}, Karoline Michalska^{1,2,3} & Andrzej Joachimiak^{1,2,3*}



Soichi Wakatsuki
SLAC NATIONAL ACCELERATOR LABORATORY

Tobias Erb
MAX PLANCK GESELLSCHAFT

The world fastest CO₂ fixation enzymes
CSP 1755

FICUS PILOT

Steve Withers
UBC

Paul Adams
ALS ADVANCED LIGHT SOURCE

Novel glycoside phosphorylase:
CSP 2572

FICUS PILOT

Andrzej Joachimiak
Argonne NATIONAL LABORATORY

Ramon Gonzalez
UNIVERSITY OF SOUTH FLORIDA

Synthetic C1-trophy Pathway
CSP 505301

FICUS PILOT

Genomes to Structure and Function Workshop



Challenges and opportunities identified through the workshop

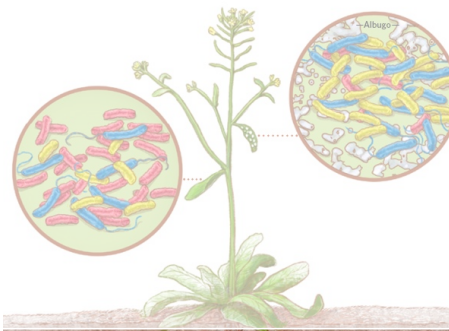
1. Science
2. Technology development
3. User Facility Integration

Overview of Research Focus Areas

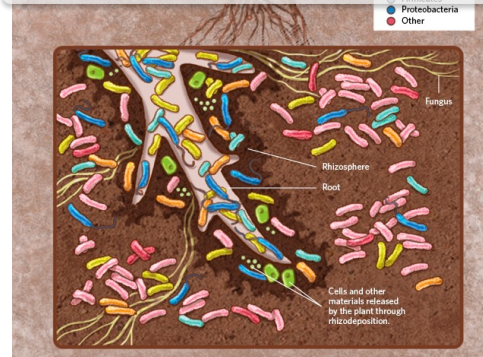


Yoshikuni group

1. Plant N-utilization



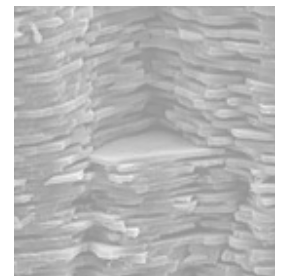
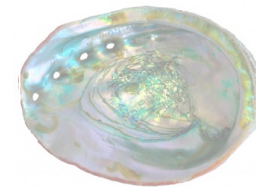
2. Microbiome engineering



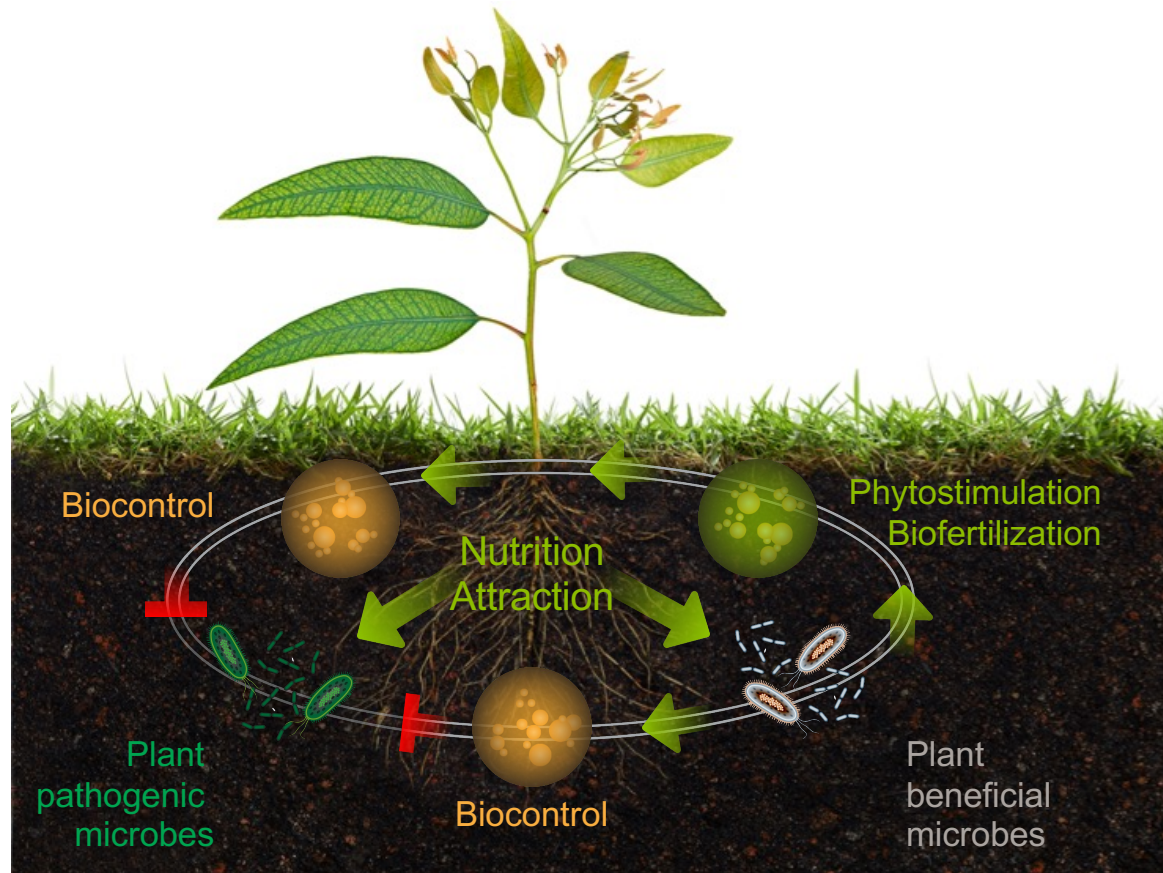
3. Biofuel and Bioproducts



4. Hybrid Biomaterials



Plant-Microbe Interactions in Sustainable Agriculture



Microbiome Engineering in Agriculture

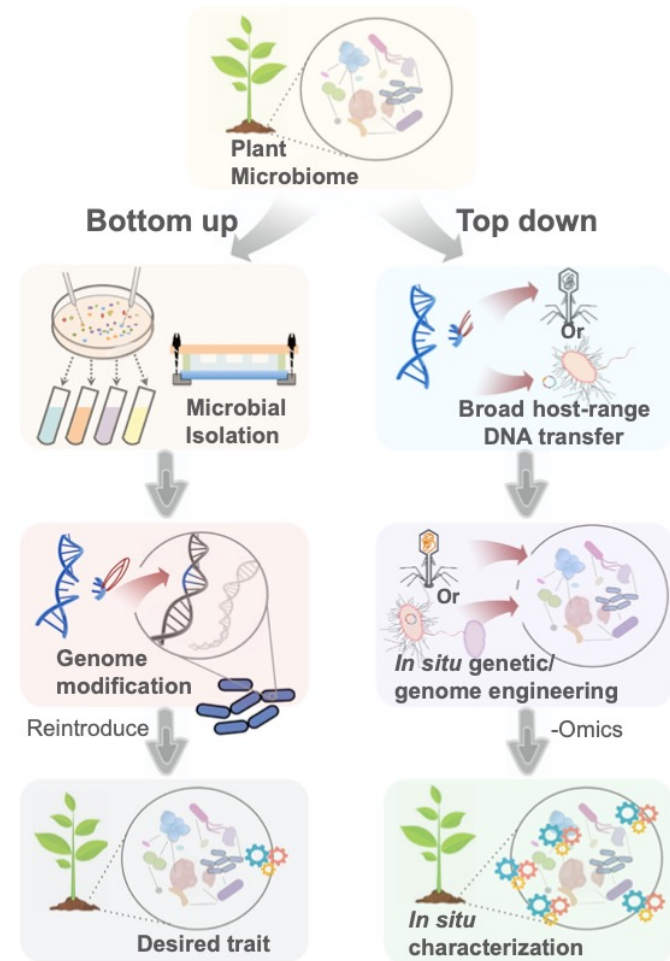
Trends in Biotechnology (2021)

Review

Microbiome Engineering: Synthetic Biology of Plant-Associated Microbiomes in Sustainable Agriculture

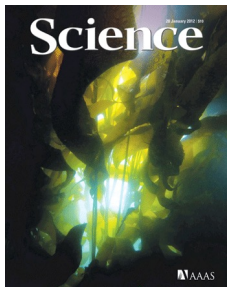
Jing Ke,^{1,6} Bing Wang,^{1,6} and Yasuo Yoshikuni^{1,2,3,4,5,*}

CellPress
REVIEWS



1. Tool Development for Microbiome Engineering

Development Of Chassis Independent Recombinase-assisted Genome Engineering (CRAGE)



An Engineered Microbial Platform for Direct Biofuel Production from Brown Macroalgae

Adam J. Wargacki,^{1*} Effendi Leonard,^{1*} Maung Nyan Win,^{1*} Drew D. Regitsky,¹ Christine Nicole S. Santos,¹ Peter B. Kim,¹ Susan R. Cooper,¹ Ryan M. Raisner,¹ Asael Herman,¹ Alicia B. Svititz,¹ Arun Lakshmanaswamy,¹ Yuki Kashiyama,^{1,2,3} David Baker,⁴ Yasuo Yoshikuni^{1,5}

ACS Synthetic Biology

CRAGE-Duet Facilitates Modular Assembly of Biological Systems for Studying Plant–Microbe Interactions

Bing Wang,¹ Zhiying Zhao,¹ Lauren K. Jabusch, Dawn M. Chiniquy, Koyo Ono, Jonathan M. Conway, Zheyun Zhang, Gaoyan Wang, David Robinson, Jan-Fang Cheng, Jeffery L. Dangl, Trent R. Northen, and Yasuo Yoshikuni*

Cite This: ACS Synth. Biol. 2020, 9, 2610–2615

Read Online



nature COMMUNICATIONS

ARTICLE

Received 24 Feb 2013 | Accepted 23 Aug 2013 | Published 23 Sep 2013

DOI: 10.1038/ncomms3503

Implementation of stable and complex biological systems through recombinase-assisted genome engineering

Christine Nicole S. Santos^{1,†}, Drew D. Regitsky^{1,†} & Yasuo Yoshikuni¹

nature protocols

PROTOCOL

Engineering complex biological systems in bacteria through recombinase-assisted genome engineering

Christine Nicole S Santos^{1,4} & Yasuo Yoshikuni^{1–3}

¹Bio Architect Lab, Inc., Berkeley, California, USA, ²BALChile S.A., Santiago, Chile, ³BAL Biofuels S.A., Santiago, Chile, ⁴Present address: Manu Biosynthesis, Inc., Cambridge, Massachusetts, USA. Correspondence should be addressed to Y.Y. (yoshikuni@ba-lab.com).

PLOS ONE

RESEARCH ARTICLE

Bacterial genome editing by coupling Cre-lox and CRISPR-Cas9 systems

Hualan Liu,^{1,2*} David S. Robinson^{1*}, Zong-Yen Wu^{1,3}, Rita Kuo¹, Yasuo Yoshikuni^{1,2,4}, Ian K. Blaby^{1,2}, Jan-Fang Cheng^{1,2*}

¹ US Department of Energy Joint Genome Institute, Berkeley, California, United States of America, ² Environmental Genomics and Systems Biology Division, Lawrence Berkeley National Laboratory, Berkeley, California, United States of America, ³ Department of Veterinary Medicine, National Chung Hsing University, Taichung, Taiwan, ROC, ⁴ Biological Systems and Engineering Division, Lawrence Berkeley National Laboratory, Berkeley, California, United States of America



Contents lists available at ScienceDirect

Metabolic Engineering

journal homepage: www.elsevier.com/locate/meteng



Development of platforms for functional characterization and production of phenazines using a multi-chassis approach via CRAGE

Jing Ke¹, Zhiying Zhao¹, Cameron R. Coates^{1,2}, Michalis Hadjithomas¹, Andrea Kufin¹, Katherine Louie¹, David Weller^{1,3}, Linda Thomashow^{1,4}, Nigel J. Mouncey^{1,5}, Trent R. Northen^{1,6}, Yasuo Yoshikuni^{1,2,3,4,5,6,*}

Cell Chemical Biology

CellPress

Resource

CRAGE-CRISPR facilitates rapid activation of secondary metabolite biosynthetic gene clusters in bacteria

Jing Ke,^{1,6} David Robinson,^{1,6} Zong-Yen Wu,¹ Andrea Kufin,¹ Katherine Louie,¹ Suzanne Kosina,¹ Trent Northen,^{1,2} Jan-Fang Cheng,^{1,2} and Yasuo Yoshikuni^{1,2,3,4,5,6,*}

ARTICLES

<https://doi.org/10.1038/s41564-019-0573-8>

nature microbiology

CRAGE enables rapid activation of biosynthetic gene clusters in undomesticated bacteria

Gaoyan Wang^{1,2}, Zhiying Zhao^{1,2}, Jing Ke^{1,2}, Yvonne Engel^{2,10}, Yi-Ming Shi^{2,10}, David Robinson¹, Kerem Bingol³, Zheyun Zhang¹, Benjamin Bowen^{1,4}, Katherine Louie¹, Bing Wang¹, Robert Evans¹, Yu Miyamoto¹, Kelly Cheng¹, Suzanne Kosina⁴, Markus De Raad⁴, Leslie Silva¹, Alicia Luhrs⁵, Andrea Lubbe⁵, David W. Hoyt³, Charles Francavilla⁵, Hiroshi Otani^{1,4}, Samuel Deutsch^{1,4,6}, Nancy M. Washton¹, Edward M. Rubin¹, Nigel J. Mouncey^{1,4}, Axel Visel^{1,4}, Trent Northen^{1,4}, Jan-Fang Cheng^{1,4}, Helge B. Bode^{1,7*} and Yasuo Yoshikuni^{1,4,6,8,9*}



Synthetic Biology, 2020, 5(1): ysaa015

doi: 10.1038/synbio.ysaa015

Advance Access Publication Date: 3 September 2020

Research Article

CRAGE-mediated insertion of fluorescent chromosomal markers for accurate and scalable measurement of co-culture dynamics in Escherichia coli

Avery J.C. Noonan¹, Yilin Qiu¹, Joe C.H. Ho², Jewel Ocampo², K.A. Vreugdenhil¹, R. Alexander Marr¹, Zhiying Zhao³, Yasuo Yoshikuni^{3,4,5,6,7}, and Steven J. Hallam^{1,2,8,9,10,11,*}

AMERICAN SOCIETY FOR MICROBIOLOGY Applied and Environmental Microbiology®

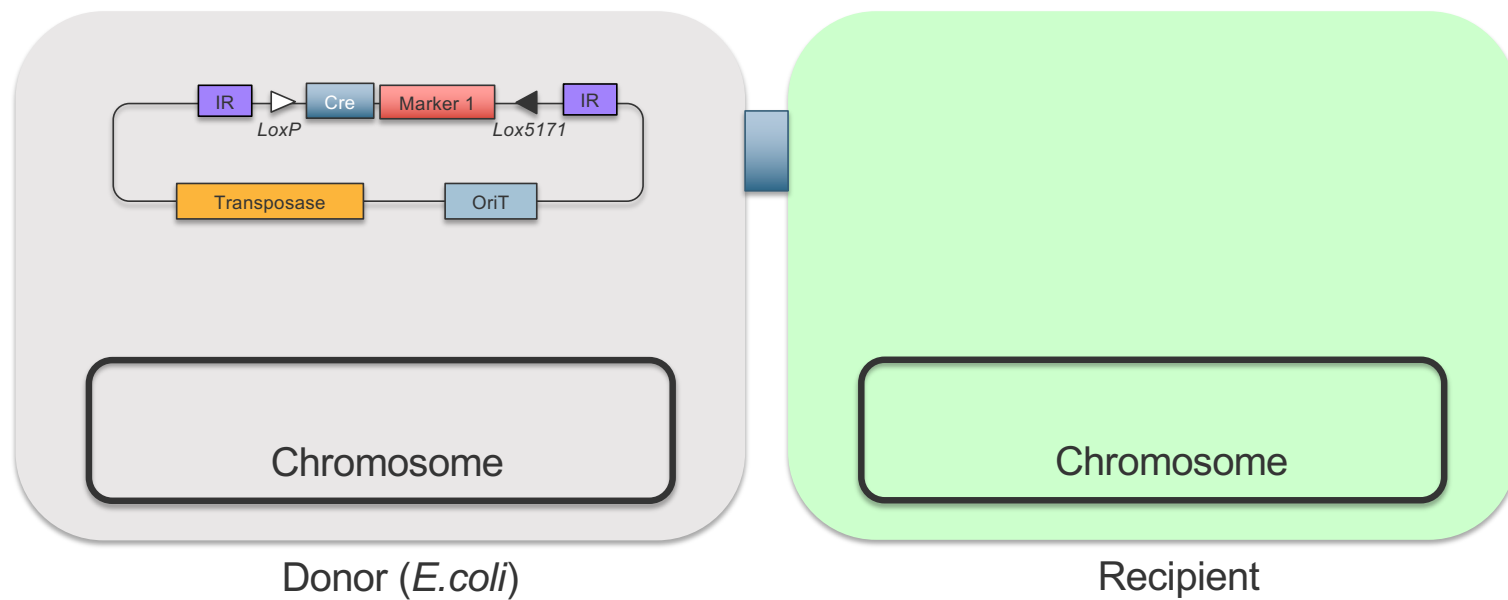
BIOTECHNOLOGY
September 2019 Volume 85 Issue 18 e01210-19
<https://doi.org/10.1128/AEM.01210-19>

Engineered Root Bacteria Release Plant-Available Phosphate from Phytate

Christine N. Shulse¹, Mansi Chovatia, Carolyn Agosto, Gaoyan Wang, Matthew Hamilton, Samuel Deutsch, Yasuo Yoshikuni, Matthew J. Blow

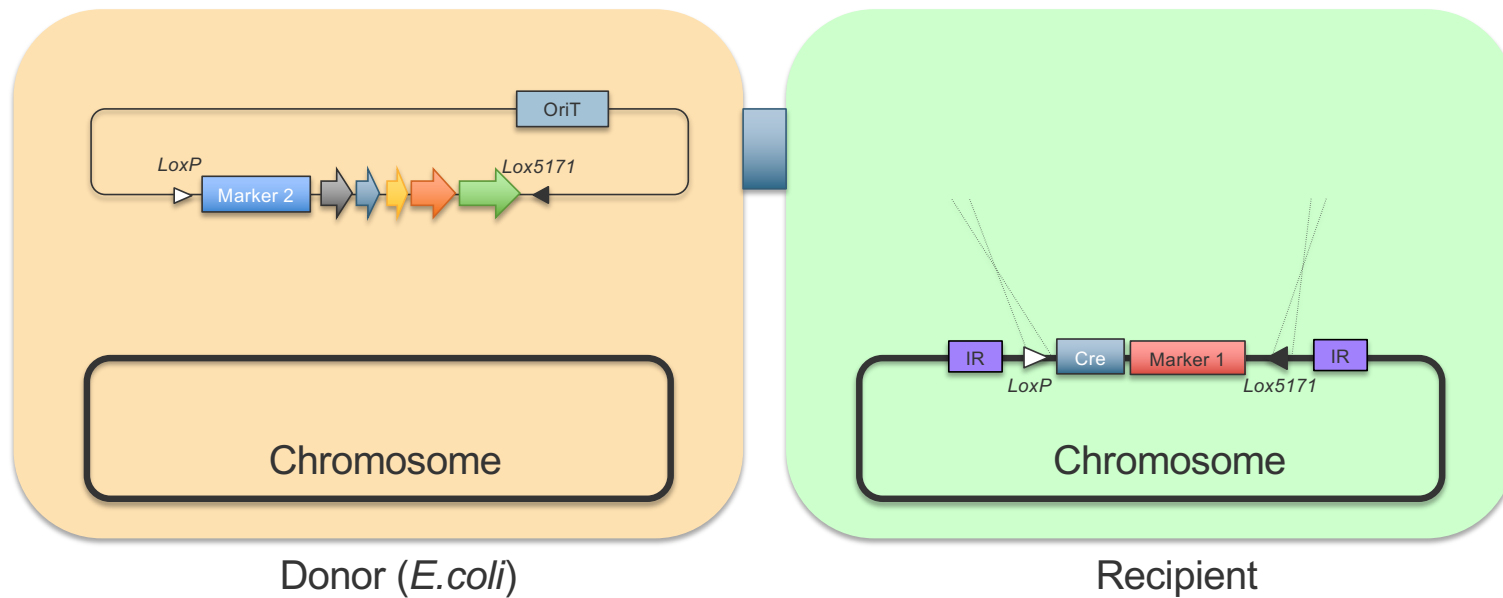
Step1: Inserting Landing Pad in Recipient Microbes

Step 1. Landing pad insertion into the genome



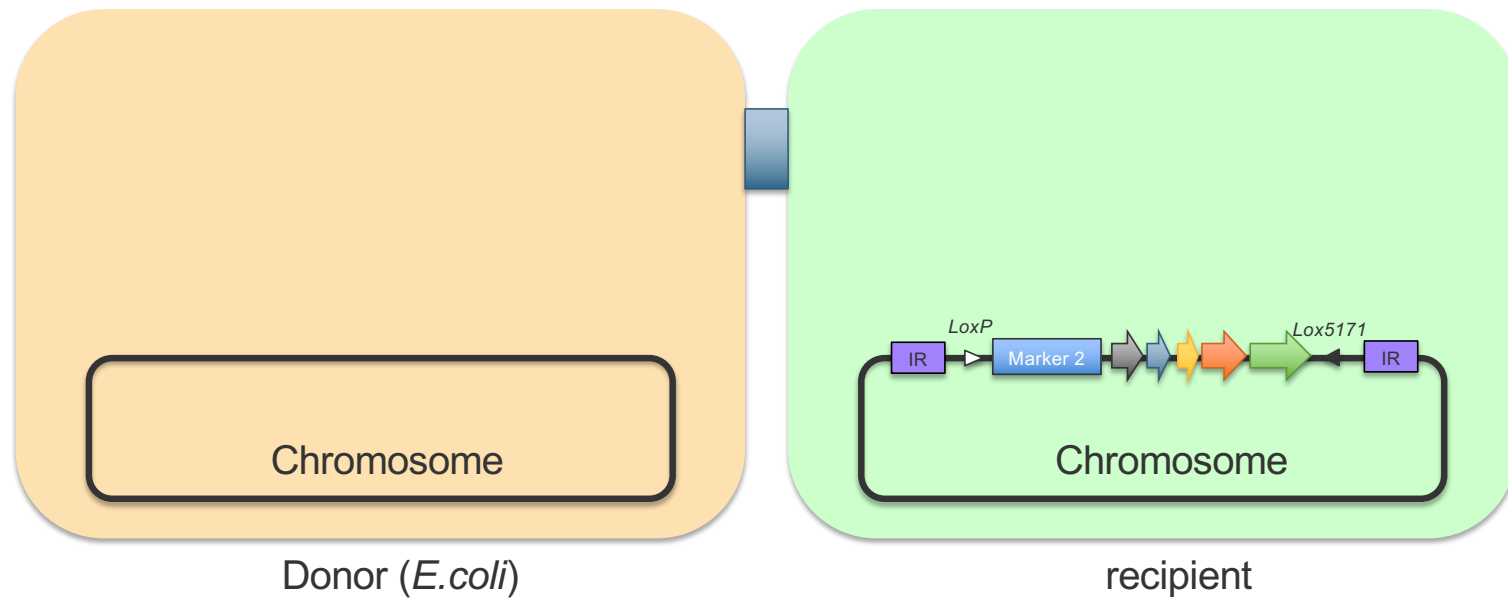
Step 2: Gene Insertion via Cre-loxP

Step 2. Gene/pathway insertion



Step 2: Gene Insertion via Cre-loxP

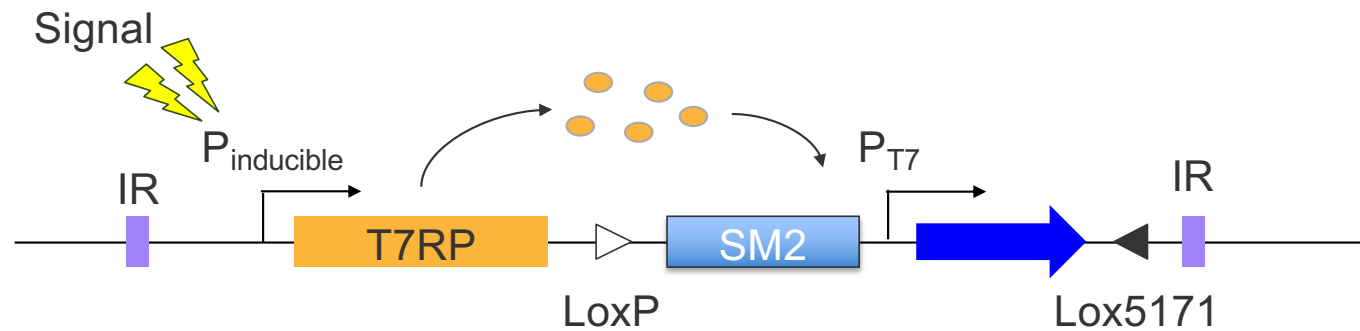
Step 2. Gene/pathway insertion



Step 3: Expression of Inserted Gene/pathway

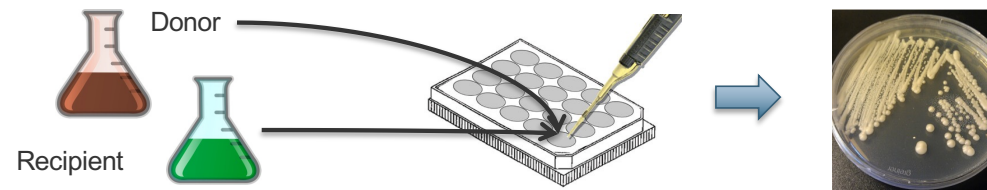
A design principle can be standardized

Step 3. Activation of inserted genes



CRAGE: Versatile Genome Engineering Tool

- Single-step pathway (~60 kbp) integration
- Transformation is automated and scaled



- Standardized design principle (a single shuttle system)
- Dual integration is feasible
- Engineering of 60+ bacterial species
 - α , β , and γ -Proteobacteria
 - Actinobacteria
 - Firmicute
 - Cyanobacteria
 - Bacteroidetes

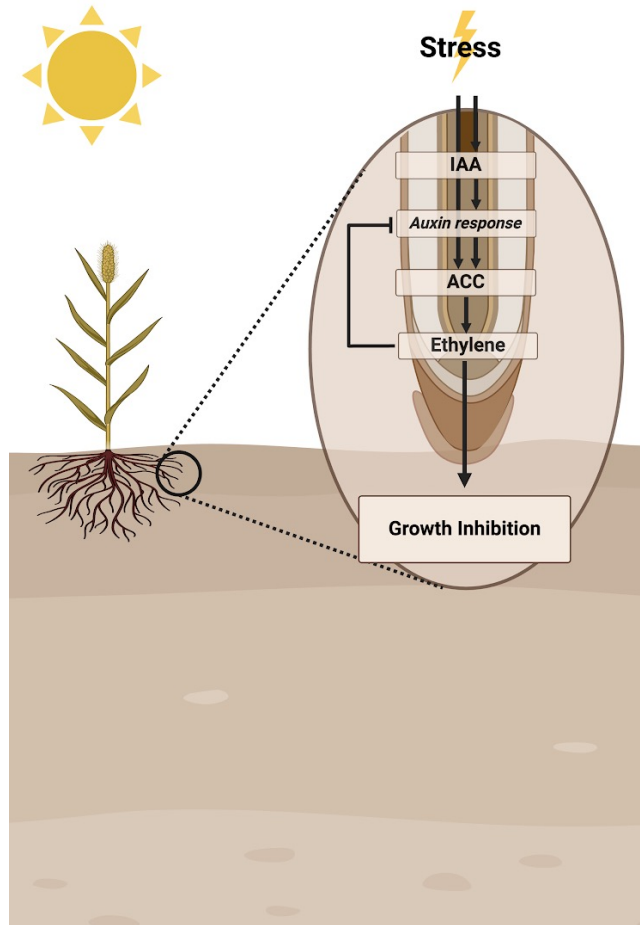
Drought Is One Of The Most Critical Issues



- **Drought impacts**

- 1.5 billion people affected by drought in 2017
- Loss of US \$125 billion globally
- Estimated impact on 75% of global population by 2050 (UNCCD, 2023)
- 23 million people severely food insecure (WFP,2023).

Microbiome Engineering Has Potential To Reduce The Negative Effect Of Drought



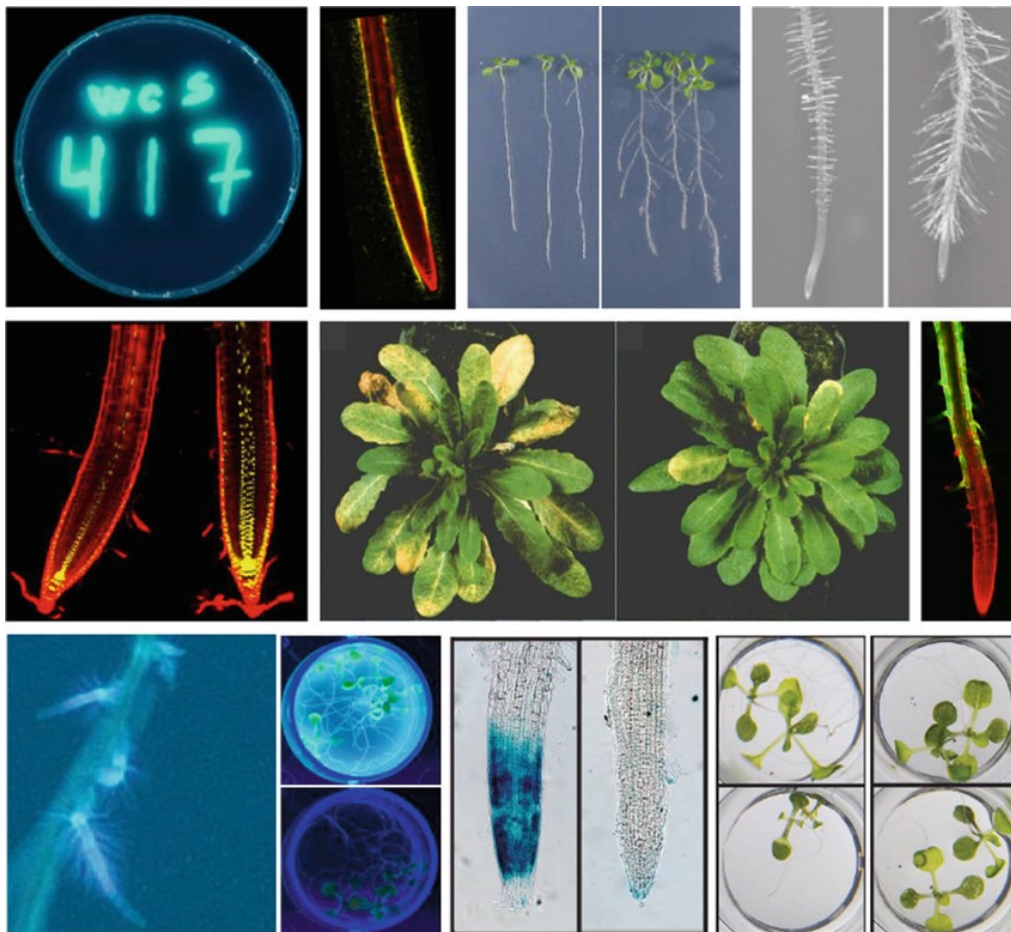
Design Principles: IAA- and AcdS-producing Syn PGPR



Chassis Selection

- 1. Persistent colonizer capable of colonizing diverse plant species**
-

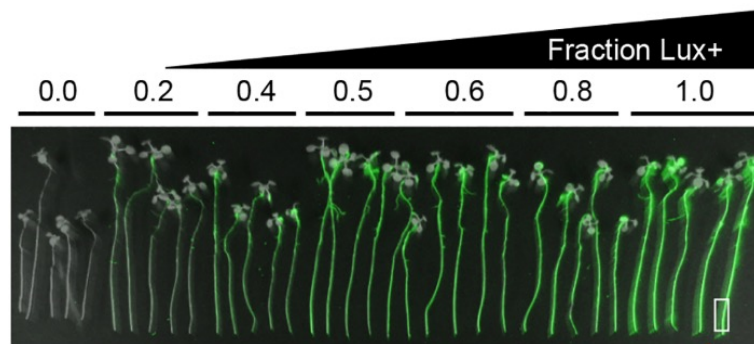
Pseudomonas simiae WCS417 is a Model PGPR



- Originally isolated from wheat fields
- Colonizes diverse plant species
- Controls soil borne diseases
 - Fungi, nematode, bacteria

P. simiae WCS417 Is A Persistent Colonizer Across The Entire Root Systems

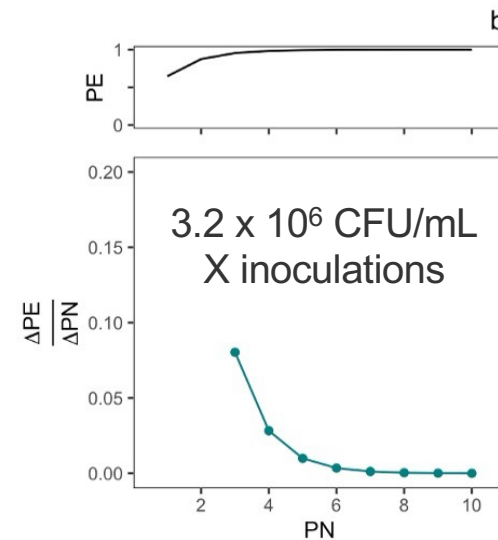
P. simiae WCS417 can colonize across the entire root systems



Root colonization of

Cole et al.
PLoS Biology (2019)

P. simiae WCS417 can persists in soil and plant roots



Klimasmith et al. Frontiers in Microbiology (in press)

Design Principles: IAA- And AcdS-producing Syn PGPR



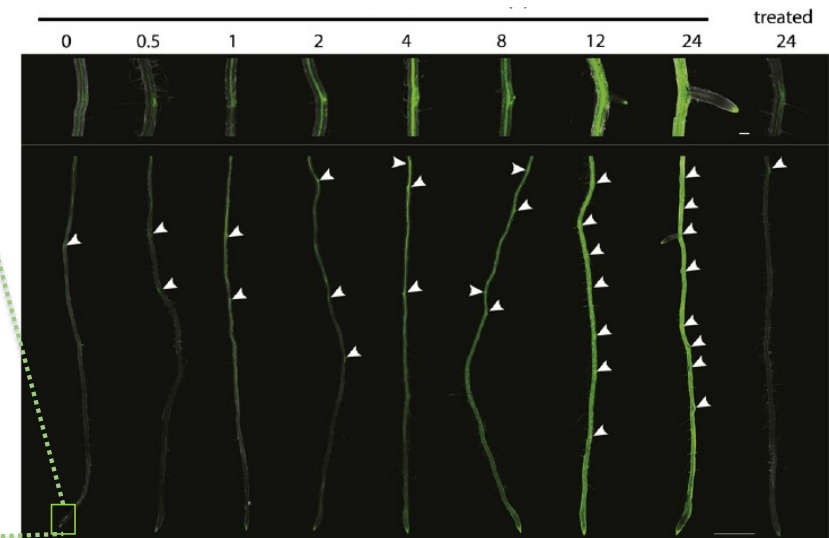
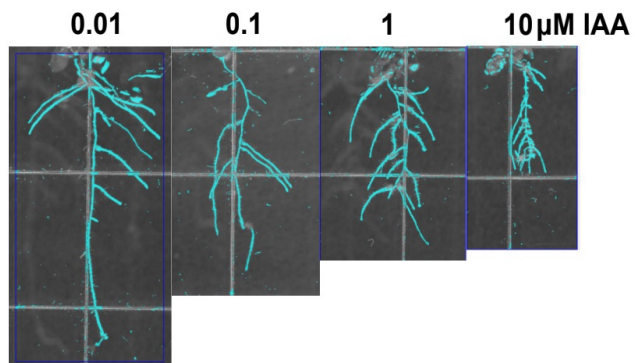
Chassis Selection

- 1. Persistent colonizer capable of colonizing diverse plant species**
 - 2. Capable of producing IAA and AcdS only around the root tips**
-

IAA Gradient Is Important For Maintaining The Plant Physiology

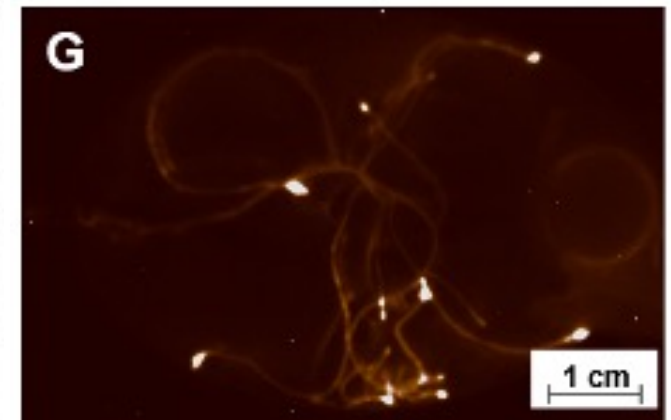
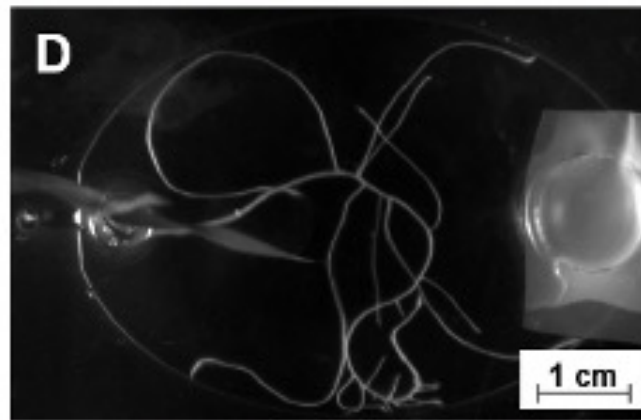
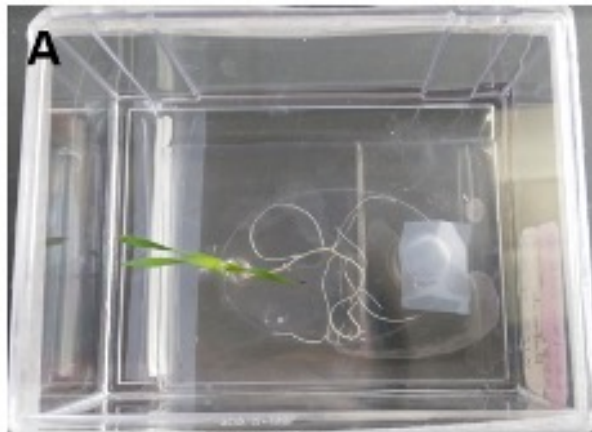
- Plant roots are sensitive to IAA
- IAA is concentrated at root tips
- Entire roots are saturated with auxin signal if treated with IAA

Auxin sensor
DR5::GFP



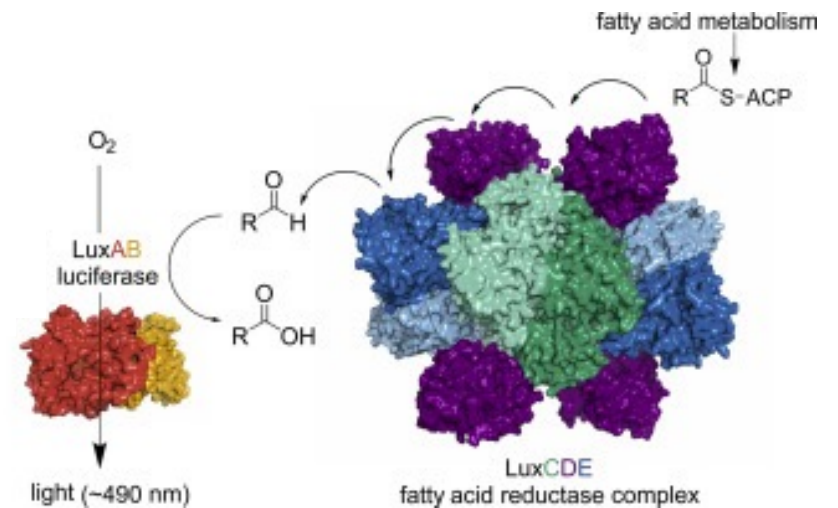
Lewis et al., *The Plant Cell* (2013) 10.1105/tpc.113.114868

P. Simiae WCS417 May Be Able To Provide IAA Activity Mainly To The Plant Root Tips



P. simiae WCS417 can colonize the root systems, but the only cells around the root tips are physiologically active

Gao et al., JoVE Journal (2018)
10.3791/57170



Design Principles: IAA- and AcdS-producing Syn PGPR



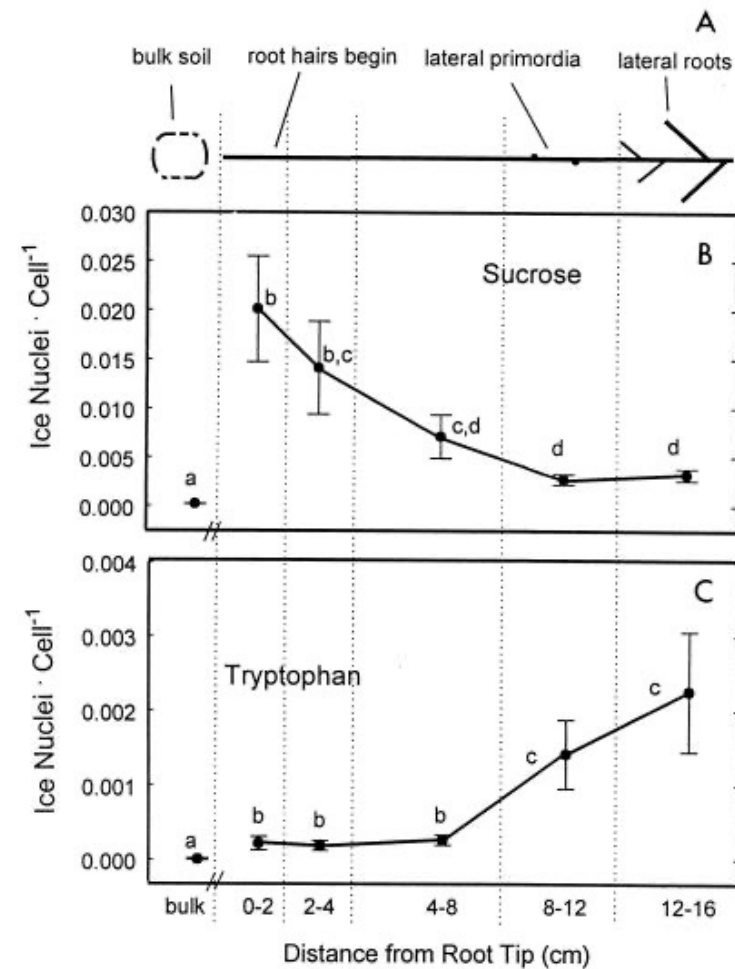
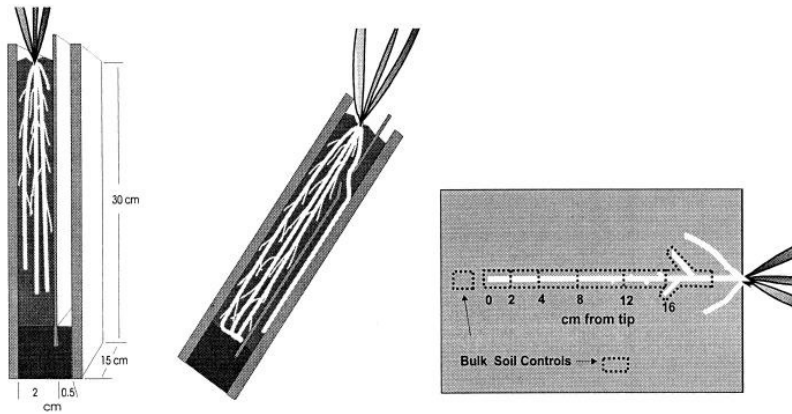
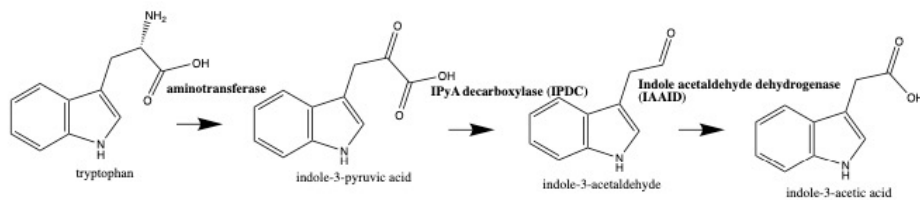
Chassis Selection

- 1. Persistent colonizer capable of colonizing diverse plant species**
- 2. Capable of producing IAA and AcdS only around the root tips**

Pathway selection

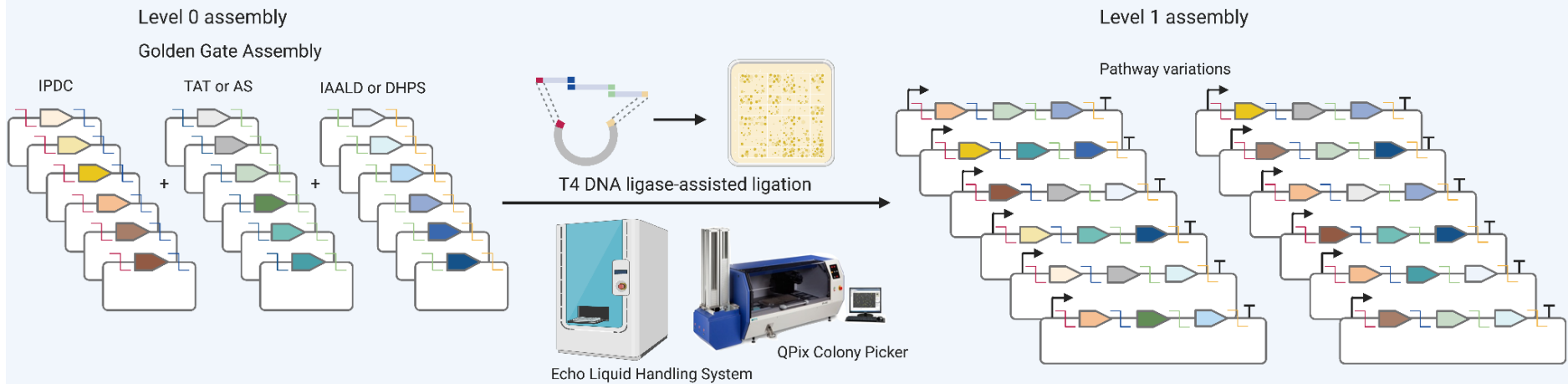
- 1. Robust IAA producer independent of Trp in plant exudates**
-

Plants Produce Trp More Abundantly From The Parts Closer To Shoot

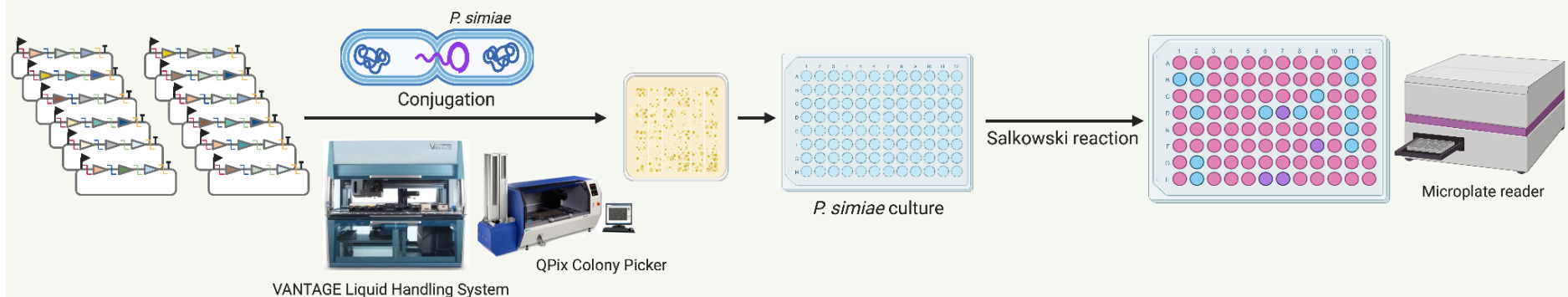


Development Of an Efficient IAA Production Strain

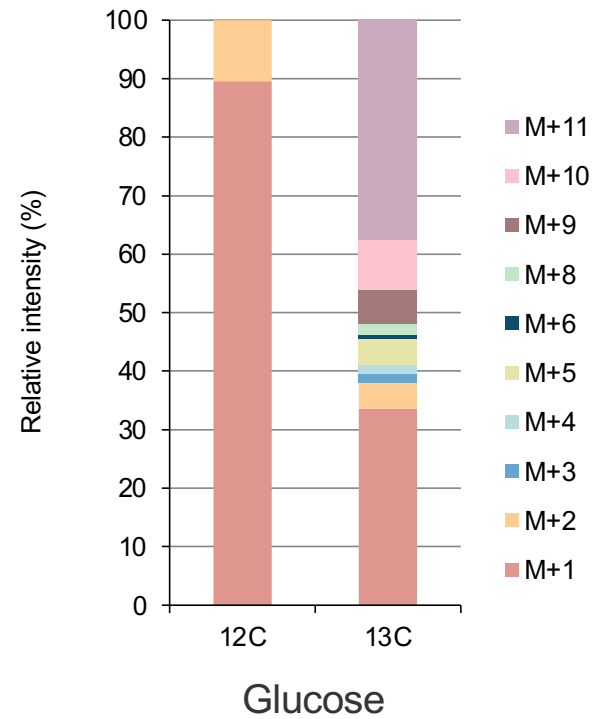
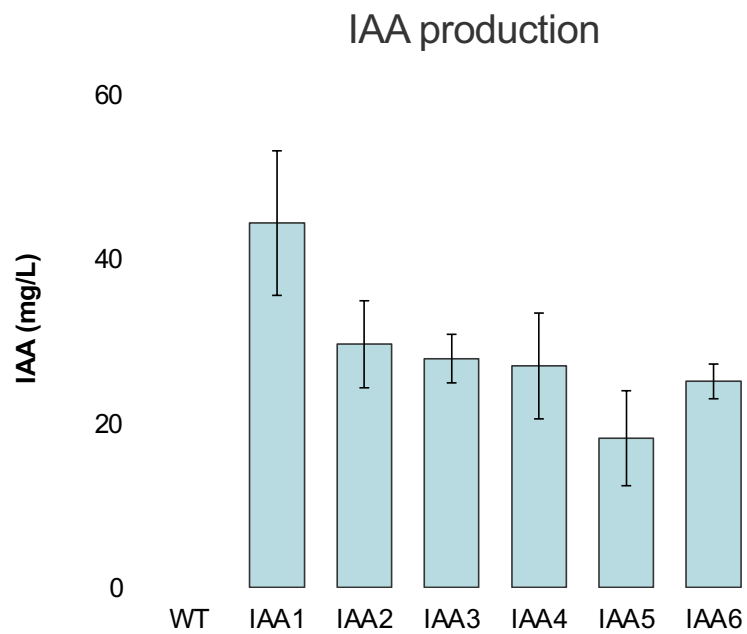
IAA biosynthesis pathway assembly



IAA biosynthesis pathway integration and IAA production measurement



Development Of An Efficient IAA Production Strain



Design Principles: IAA- And AcdS-producing Syn PGPR



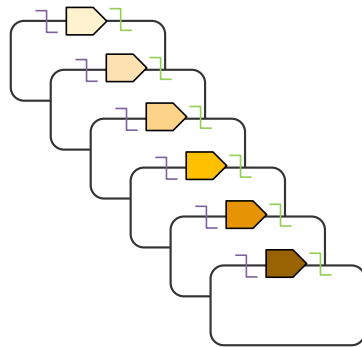
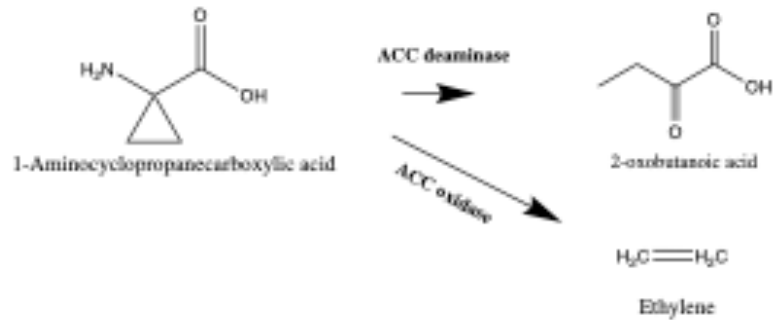
Chassis Selection

- 1. Persistent colonizer capable of colonizing diverse plant species**
- 2. Capable of producing IAA and AcdS only around the root tips**

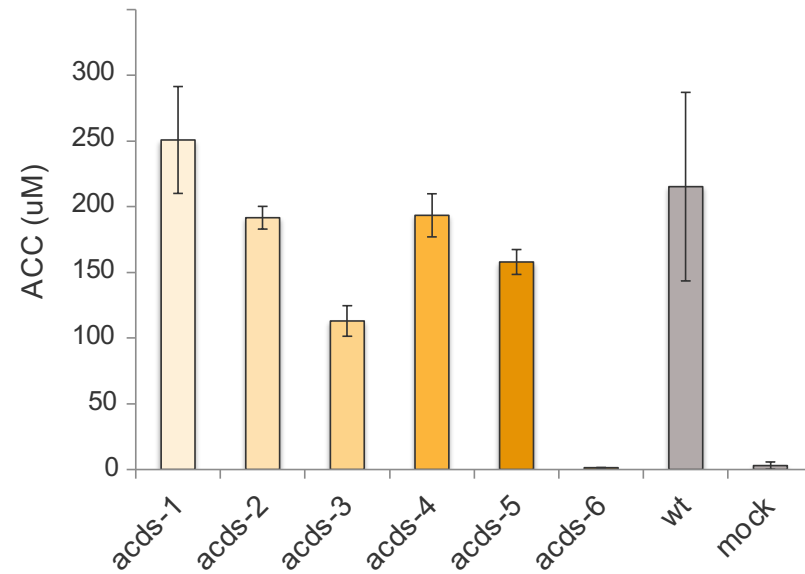
Pathway selection

- 1. Robust IAA producer independent of Trp in plant exudates**
 - 2. Robust AcdS producer**
-

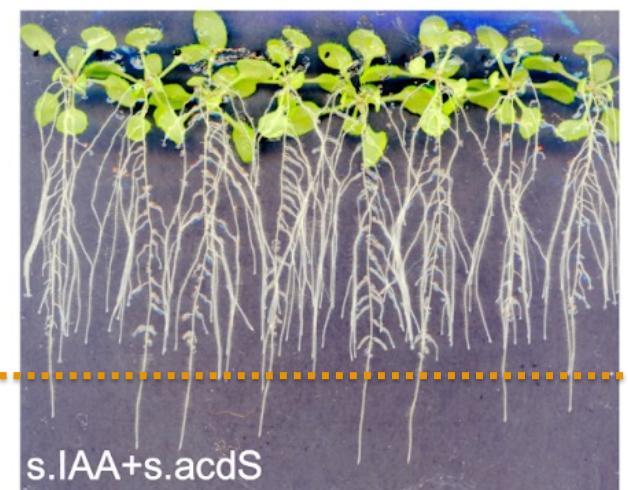
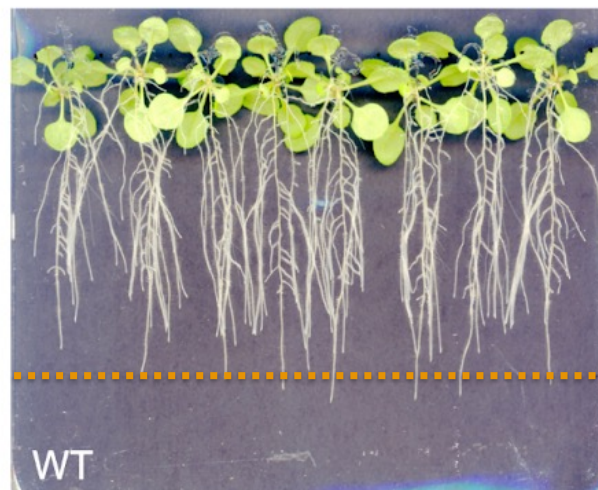
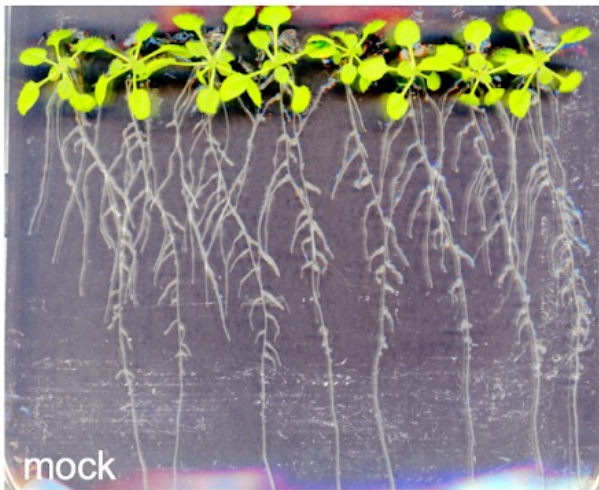
Development Of An Efficient ACC Deaminating Strain



ACC degradation

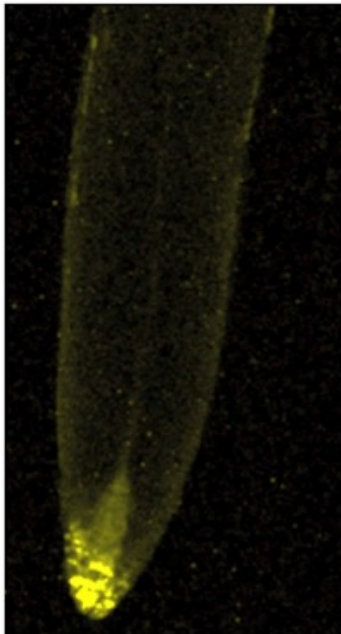


IAA- and AcdS-producing SynPGPR Can Promote The Root Growth

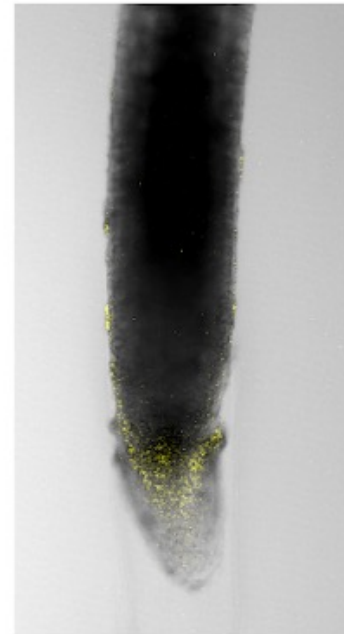
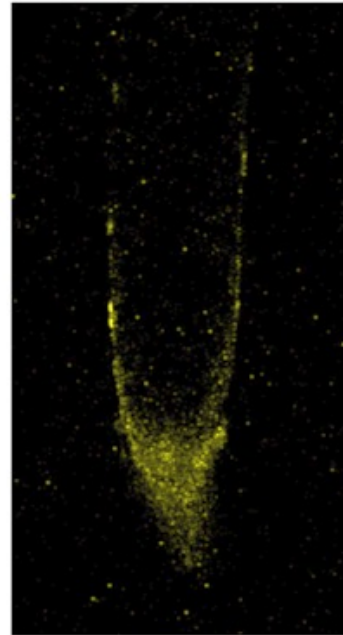


Our IAA Production Strain Can Maintain The IAA Gradient

WT

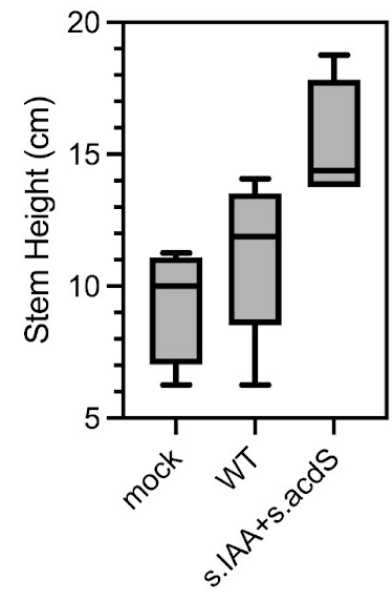
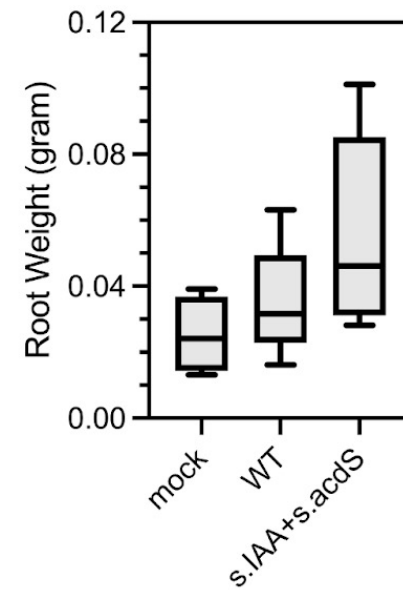
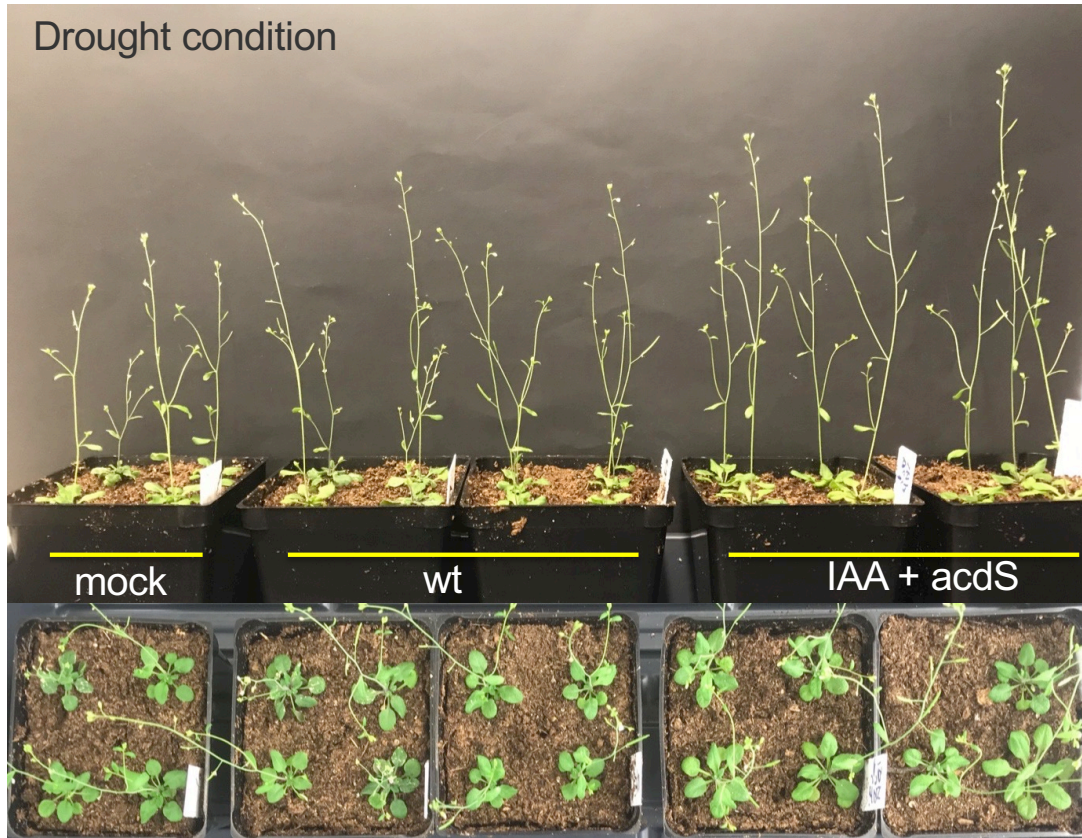


s.IAA



Our SynPGPR Can Promote The Growth Of Arabidopsis

Drought condition



Summary

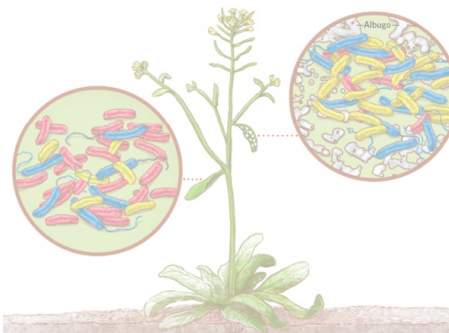
- **CRAGE is a powerful strain engineering technology for microbial engineering**
- **Engineered microbes can tremendously increase plant biomass yield under the normal and drought conditions**

Overview of Research Focus Areas

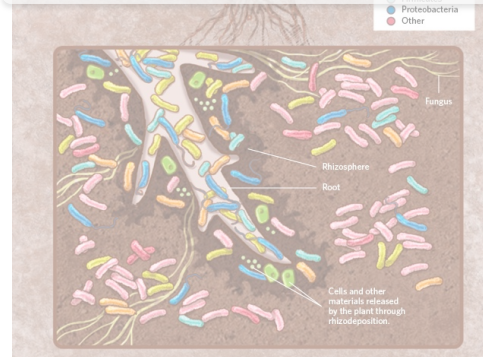


Yoshikuni group

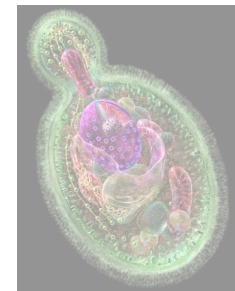
1. Plant N-utilization



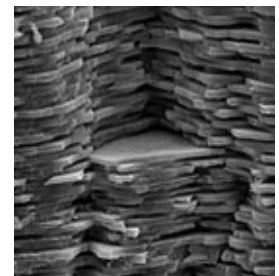
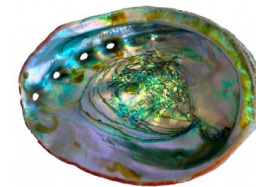
2. Microbiome engineering



3. Biofuel and Bioproducts



4. Hybrid Biomaterials

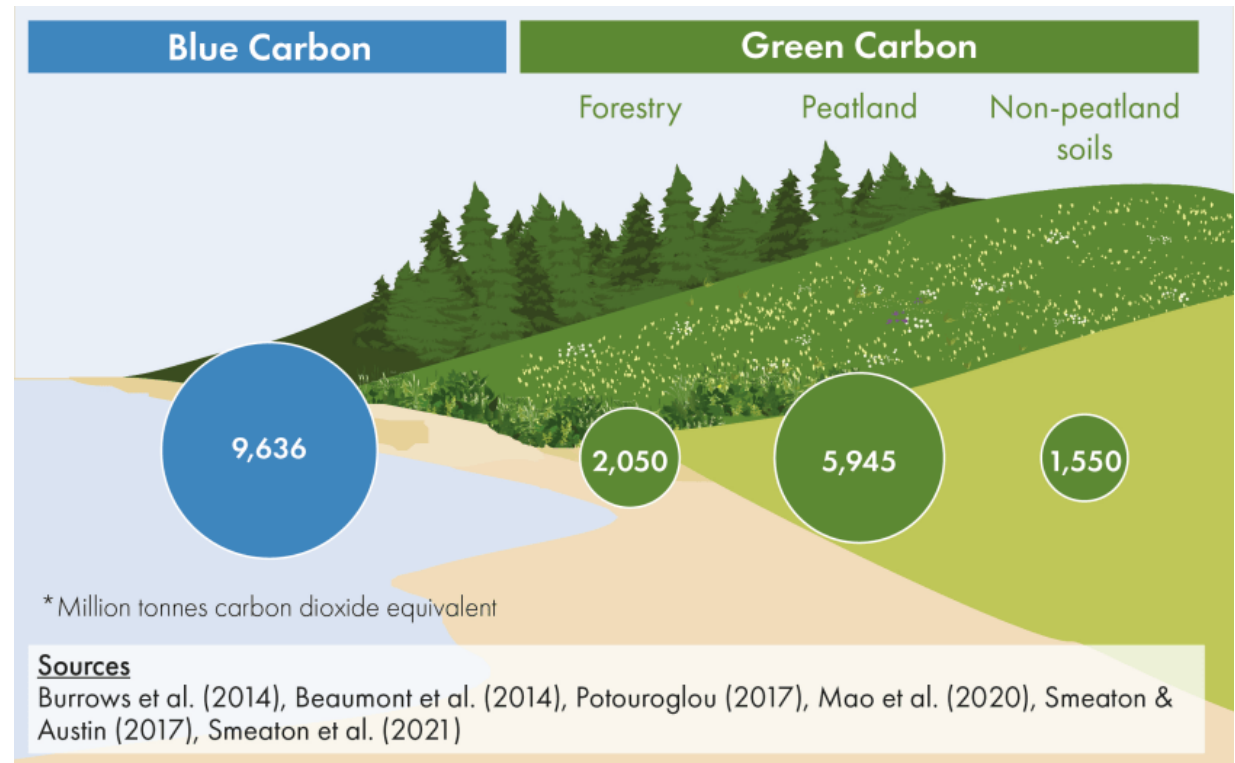


Carbons with Different Colors



BLACK CARBON

Carbon particles given off by hot fires, like coal plants, forest fires, and combustion from cars



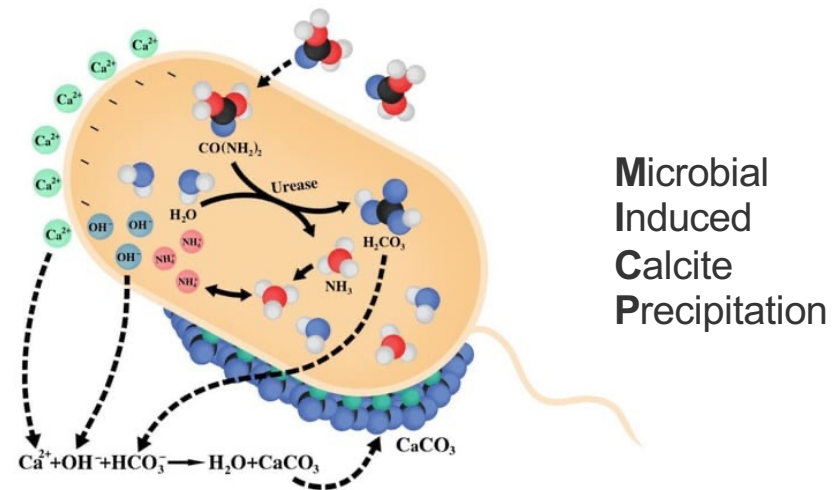
A new carbon with different color – yellow?



- N, P, and K rather than C
 - 2.8 kg-CO_{2eq}/kg-NH₃
 - 1.9 kg-CO_{2eq}/kg-P₂O₅
 - 0.5 kg-CO_{2eq}/kg-K₂O
- Urine composes only 1% of total wastewater, but it contains roughly 70-90% of nitrogen (N) and 50-65% of phosphorus (P) in waste streams
- **Urine diversion process** - If we can fully utilize urine, 21%, 12%, and 20% of global N-, P-, and K-fertilizer demands are met
- Problem: low price of fertilizers

What chemicals should we make to subsidize urine diversion system

Chemical	Concentration in g/100 ml urine
Water	95
Urea	2
Sodium	0.6
Chloride	0.6
Sulfate	0.18
Potassium	0.15
Phosphate	0.12
Creatinine	0.1
Ammonia	0.05
Uric acid	0.03
Calcium	0.015
Magnesium	0.01
Protein	--
Glucose	--

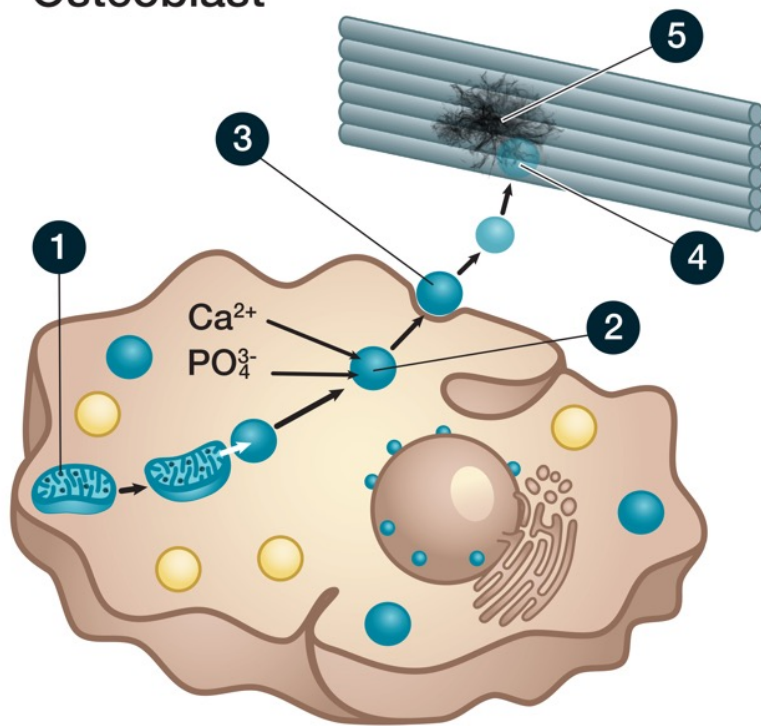


**Microbial
Induced
Calcite
Precipitation**

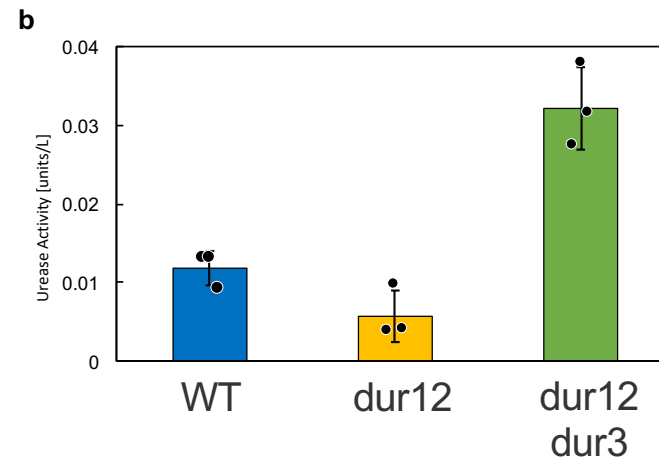
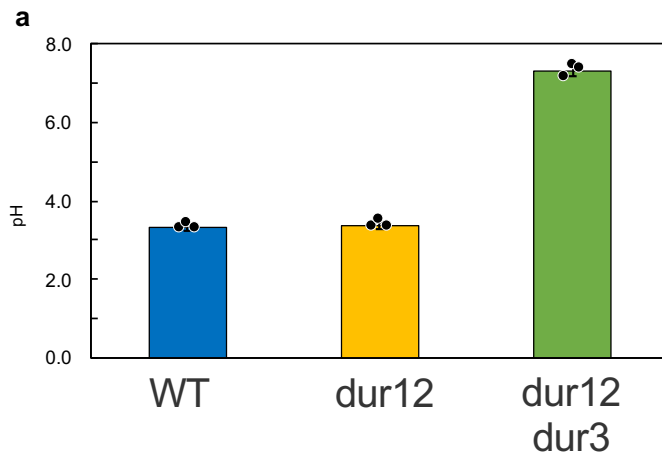
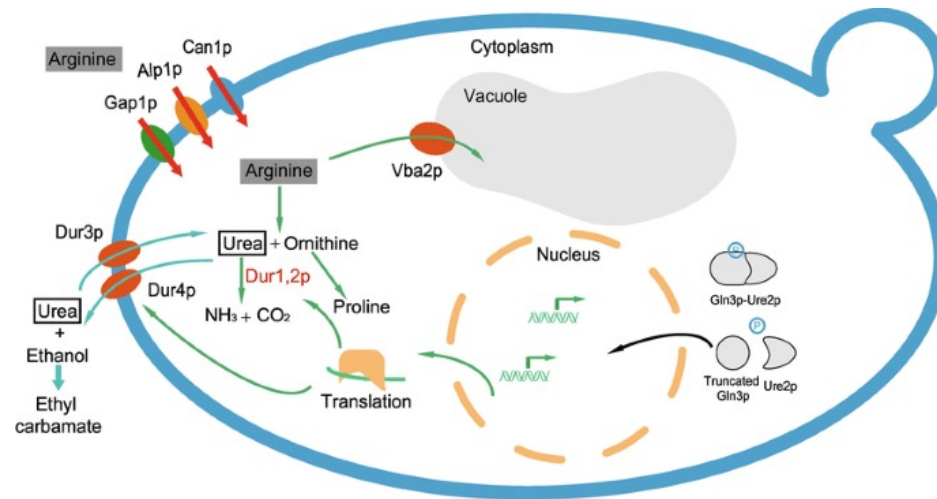
- **Hydroxyapatite might be a very interesting material to make**
 - High price point: \$80/kg - \$10,000/kg
 - \$3.1B market size
 - Orthopedic, dental care products, plastic surgery, food, and pharmaceutical applications
 - Filter, Insulation, replacement of plastics, construction materials

Design principles of osteoyeast platform

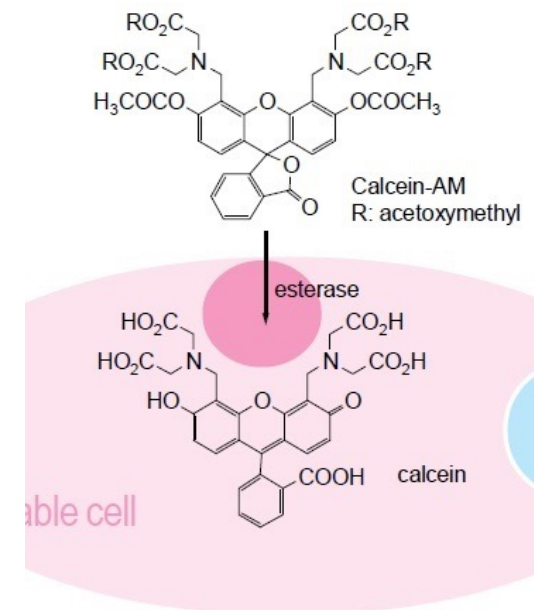
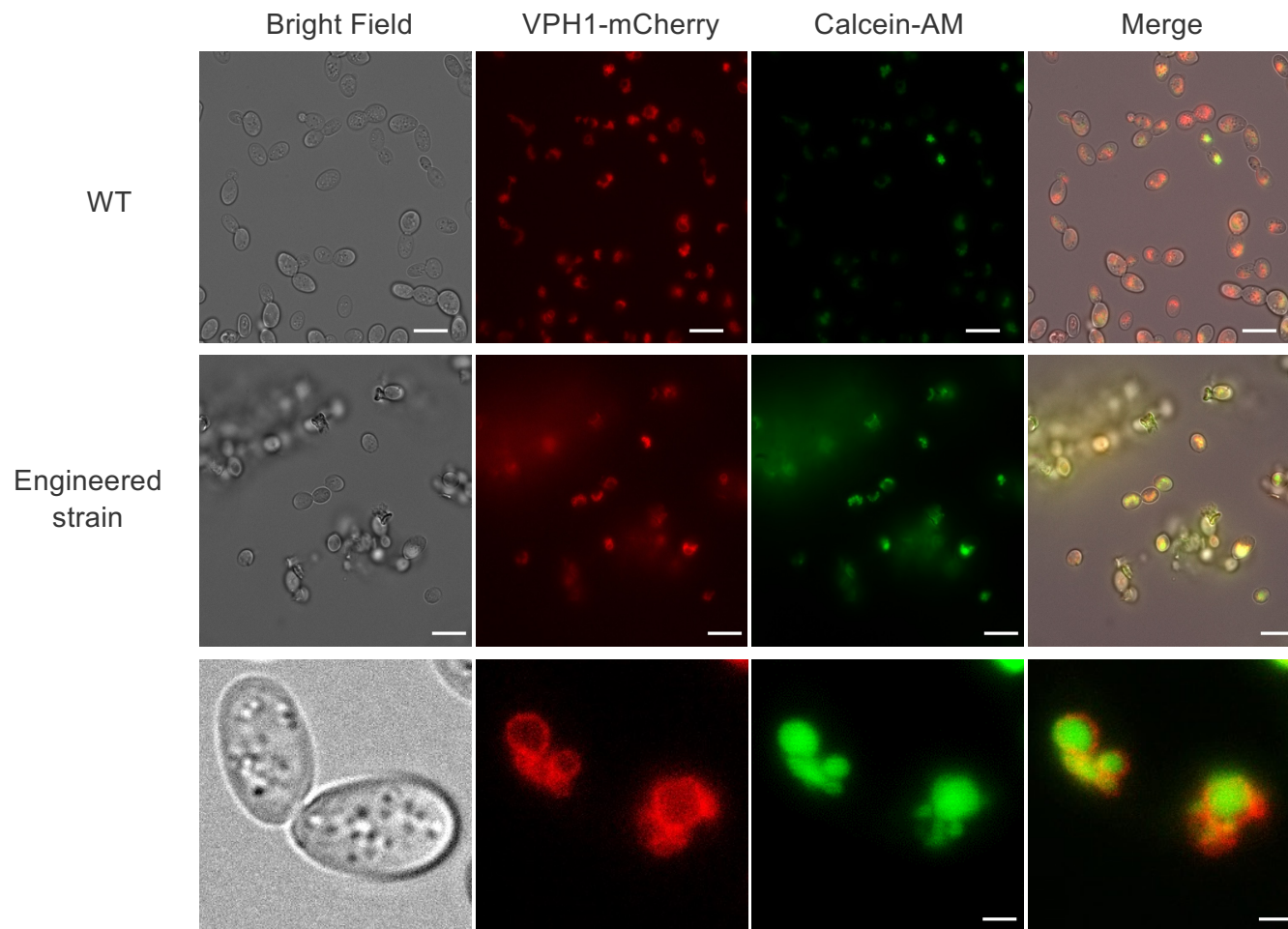
Osteoblast



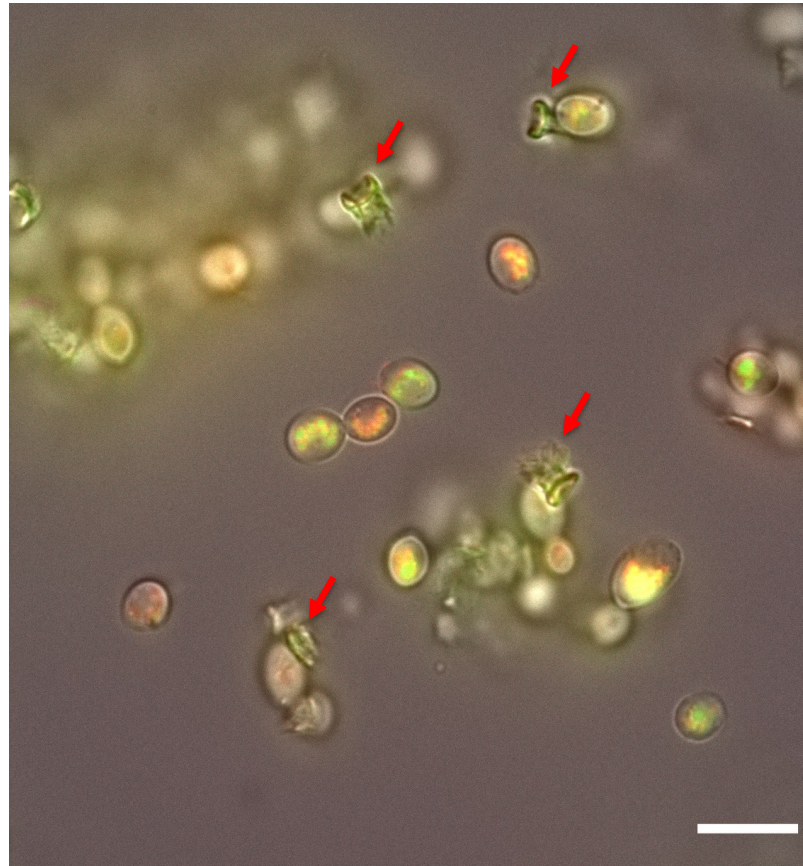
Overexpression of urea degrading enzymes



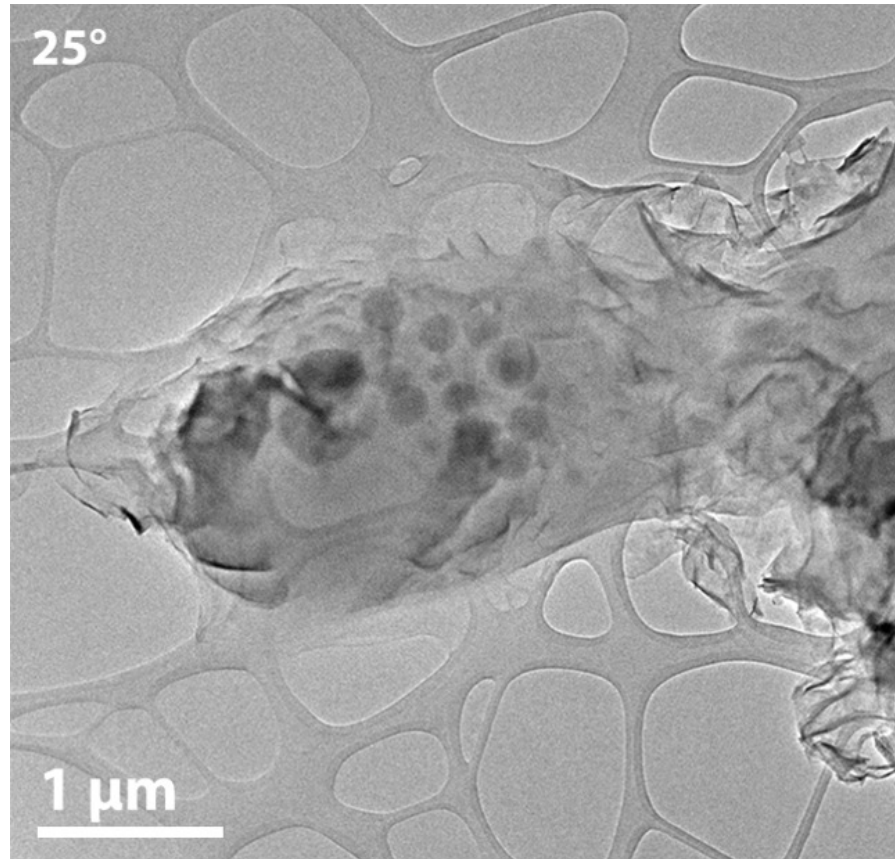
Engineered yeast increased accumulation of calcium in vacuoles



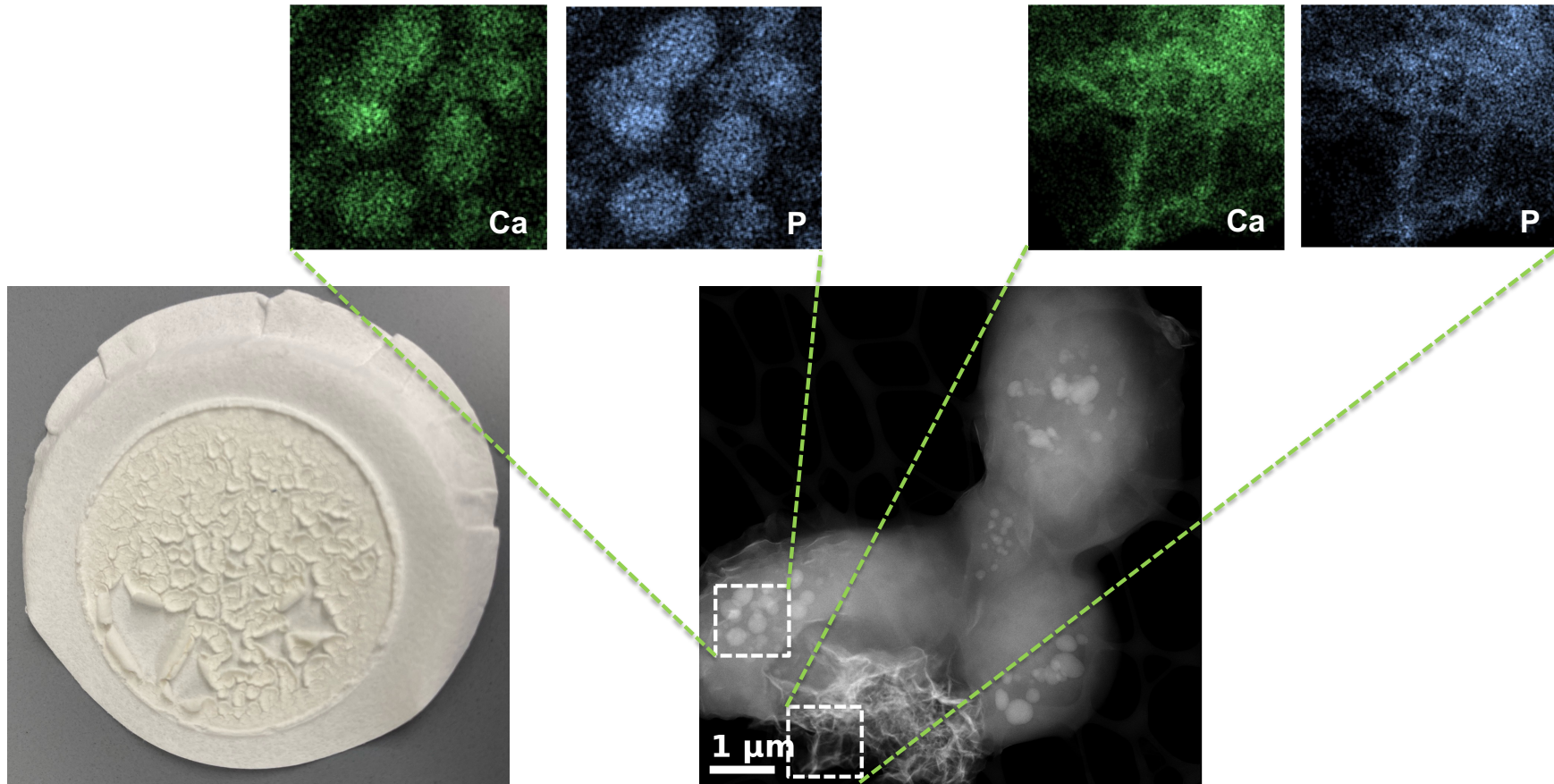
Engineered yeast produced crystals



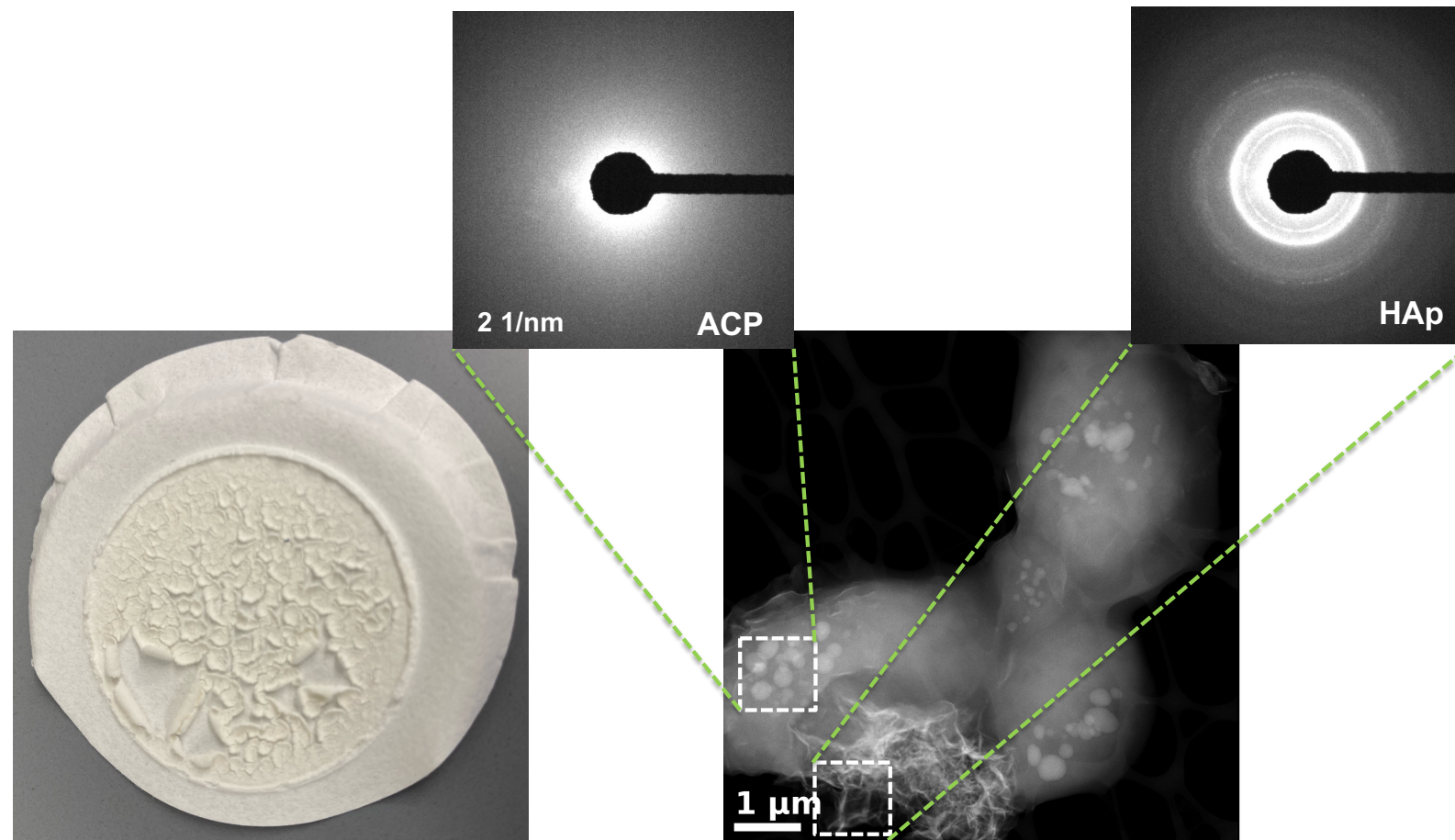
Hydroxyapatite synthesis mediated by the osteoyeast platform analyzed using TEM



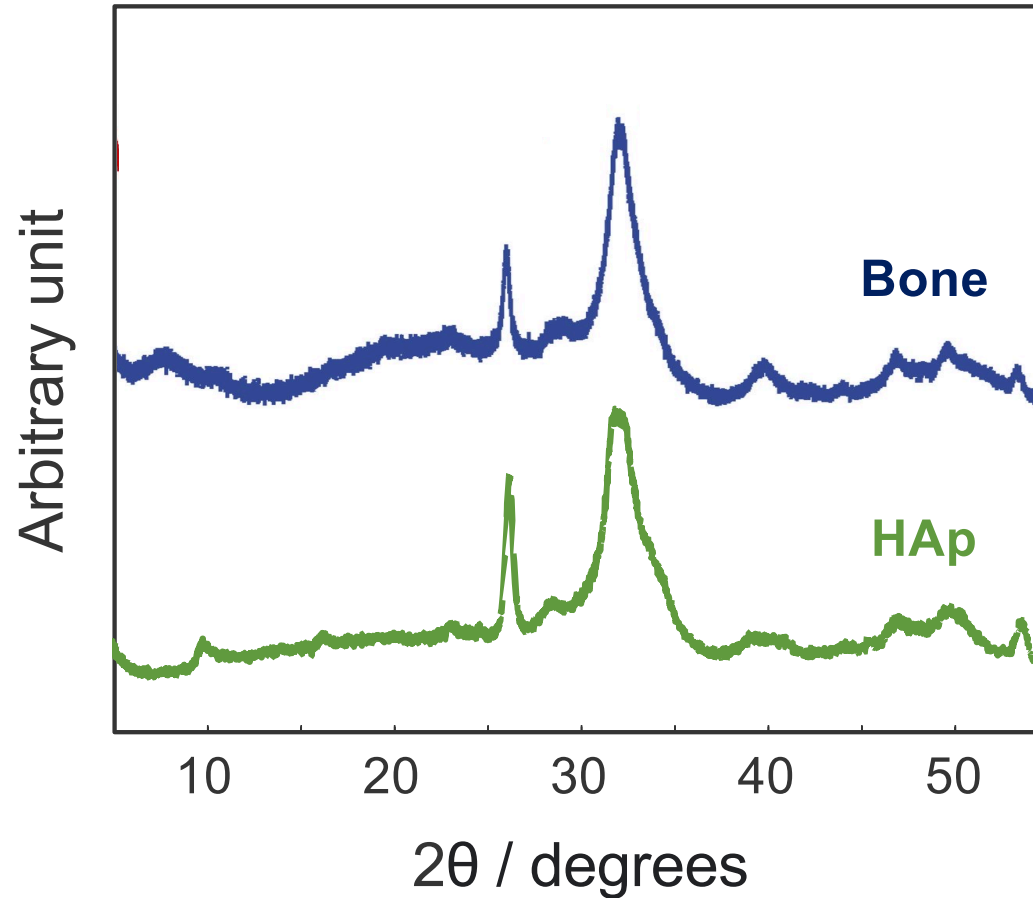
Hydroxyapatite synthesis mediated by the osteoyeast platform analyzed using TEM



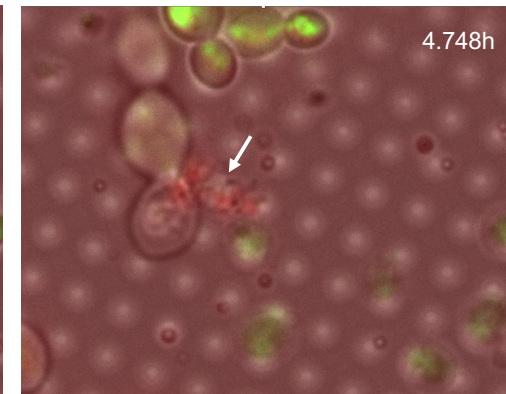
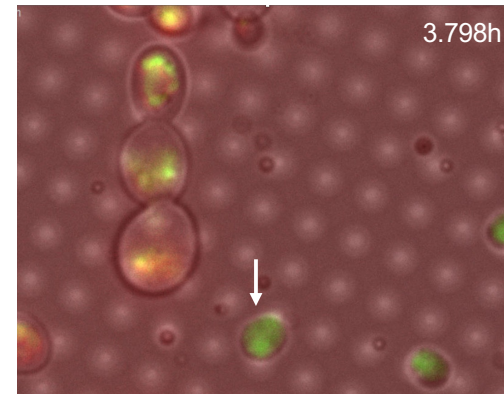
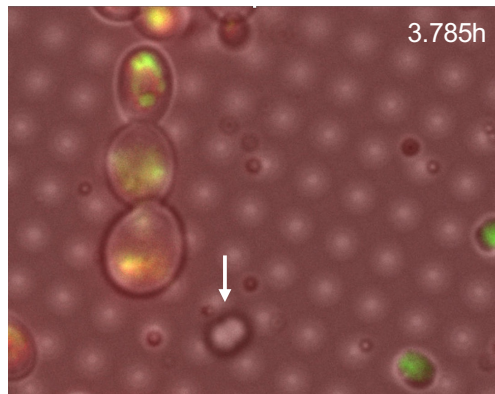
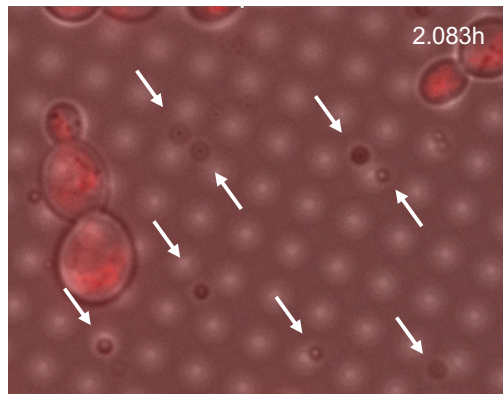
Hydroxyapatite (HAp) synthesis mediated by the osteoyeast platform analyzed using TEM



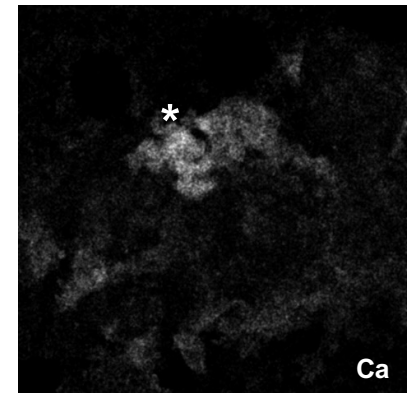
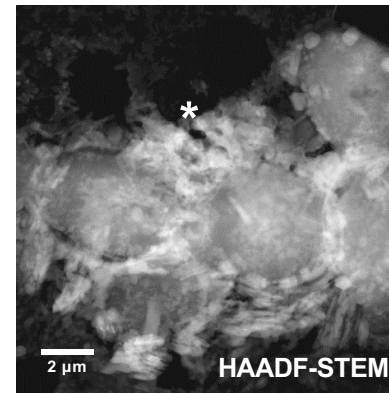
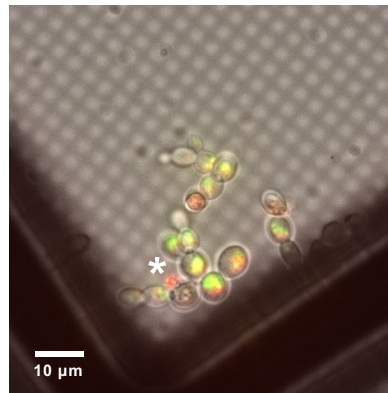
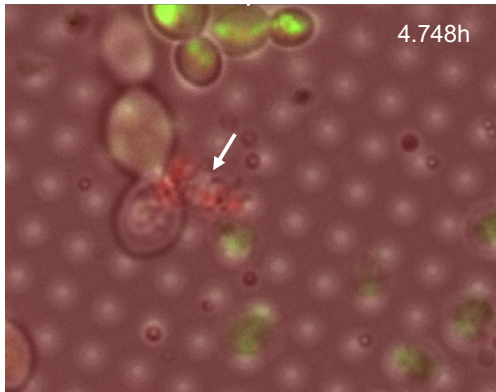
Hydroxyapatite (HAp) synthesis mediated by the osteoyeast platform analyzed using TEM



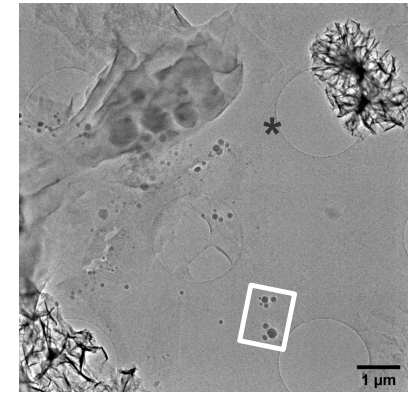
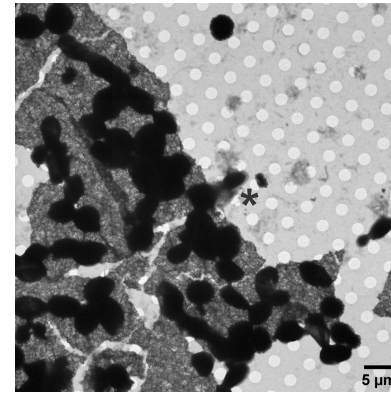
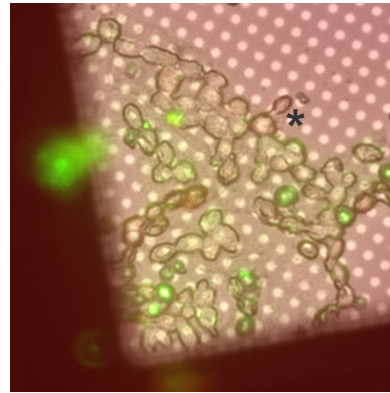
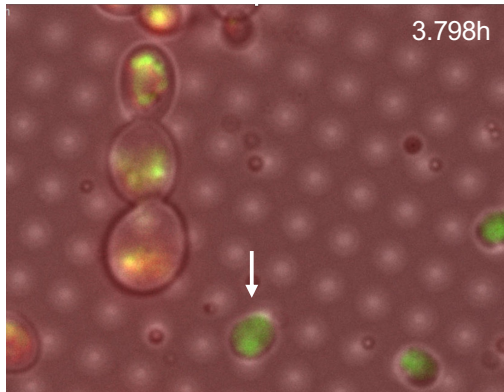
HAp synthesis mediated by the osteoyeast platform using correlative imaging analyses



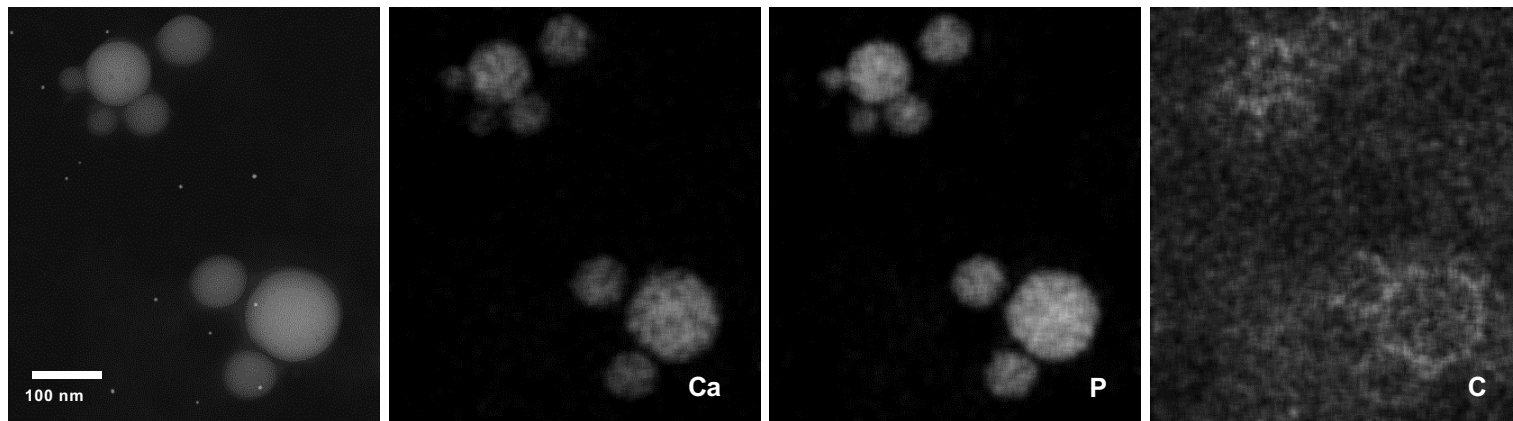
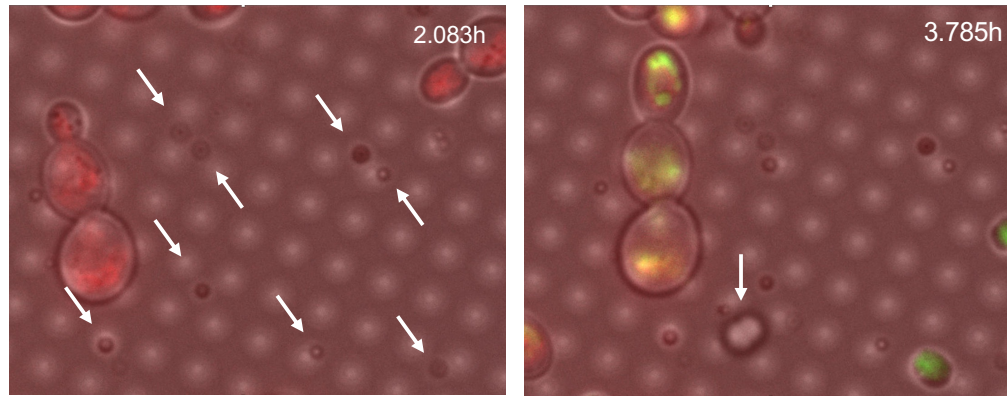
HAp synthesis mediated by the osteoyeast platform using correlative imaging analyses



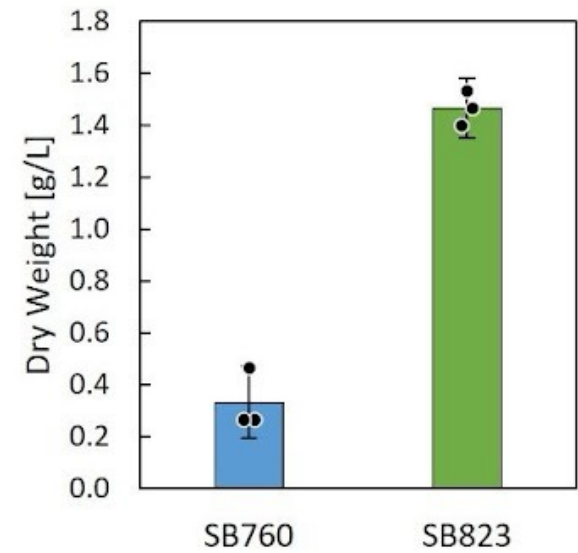
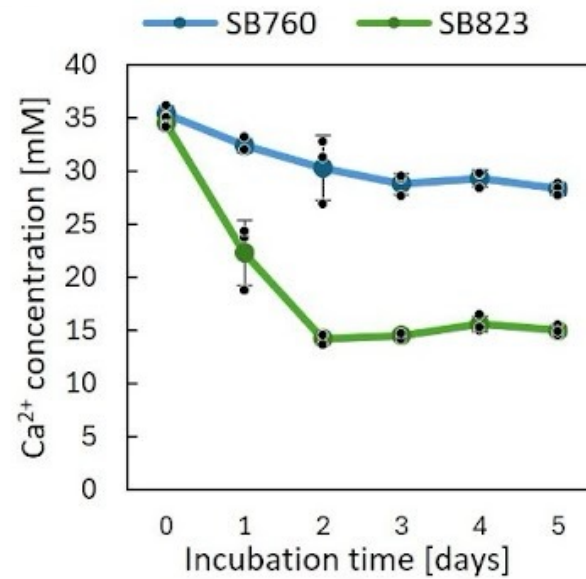
HAp synthesis mediated by the osteoyeast platform using correlative imaging analyses



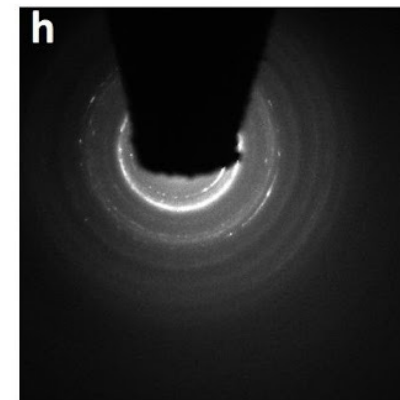
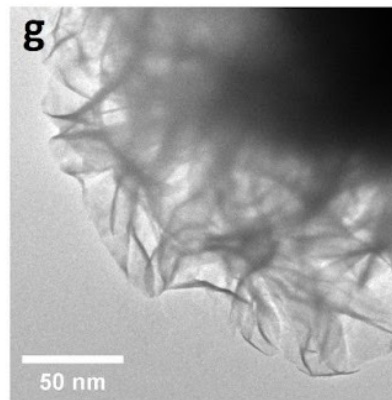
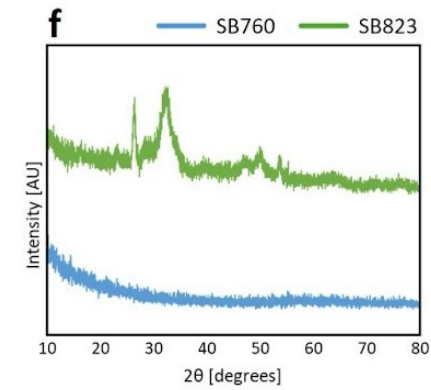
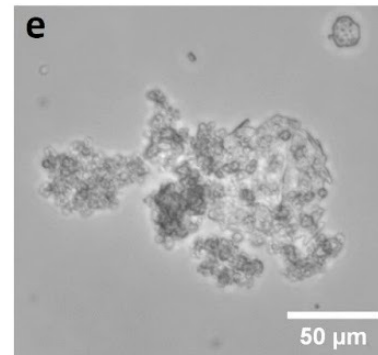
HAp synthesis mediated by the osteoyeast platform using correlative imaging analyses



HAp synthesis directly from urine

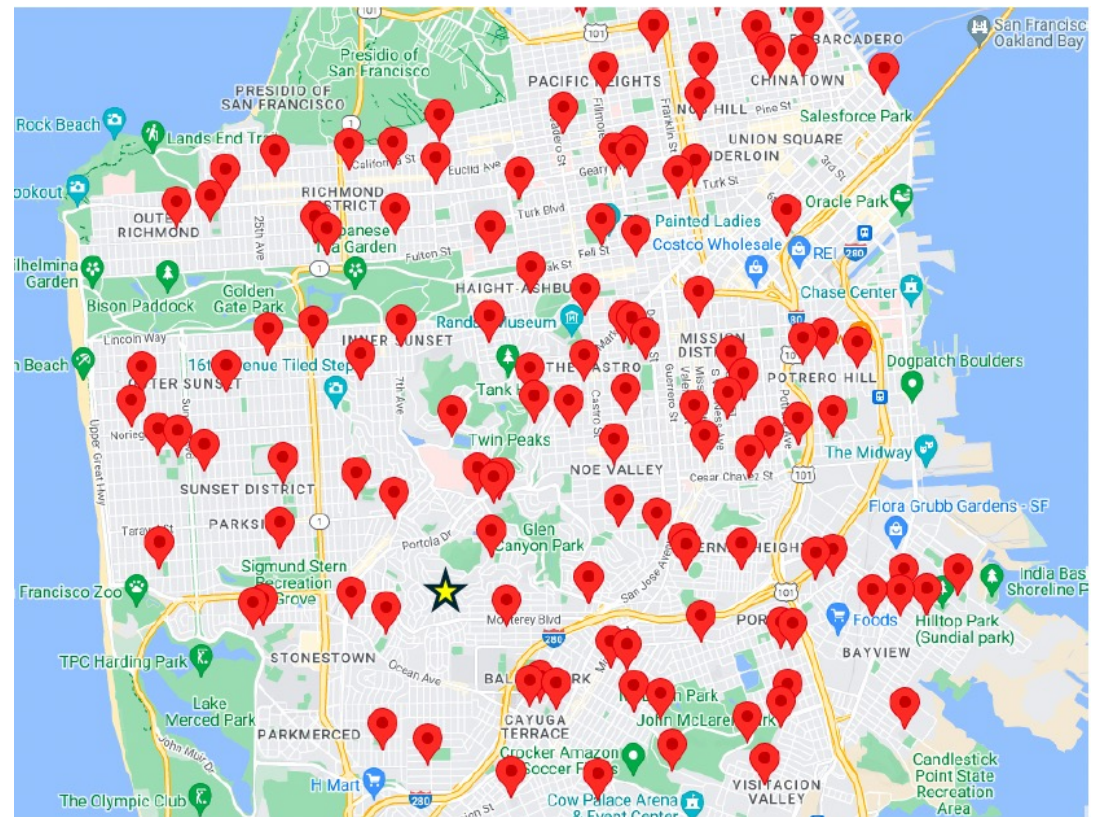


HAp synthesis directly from urine



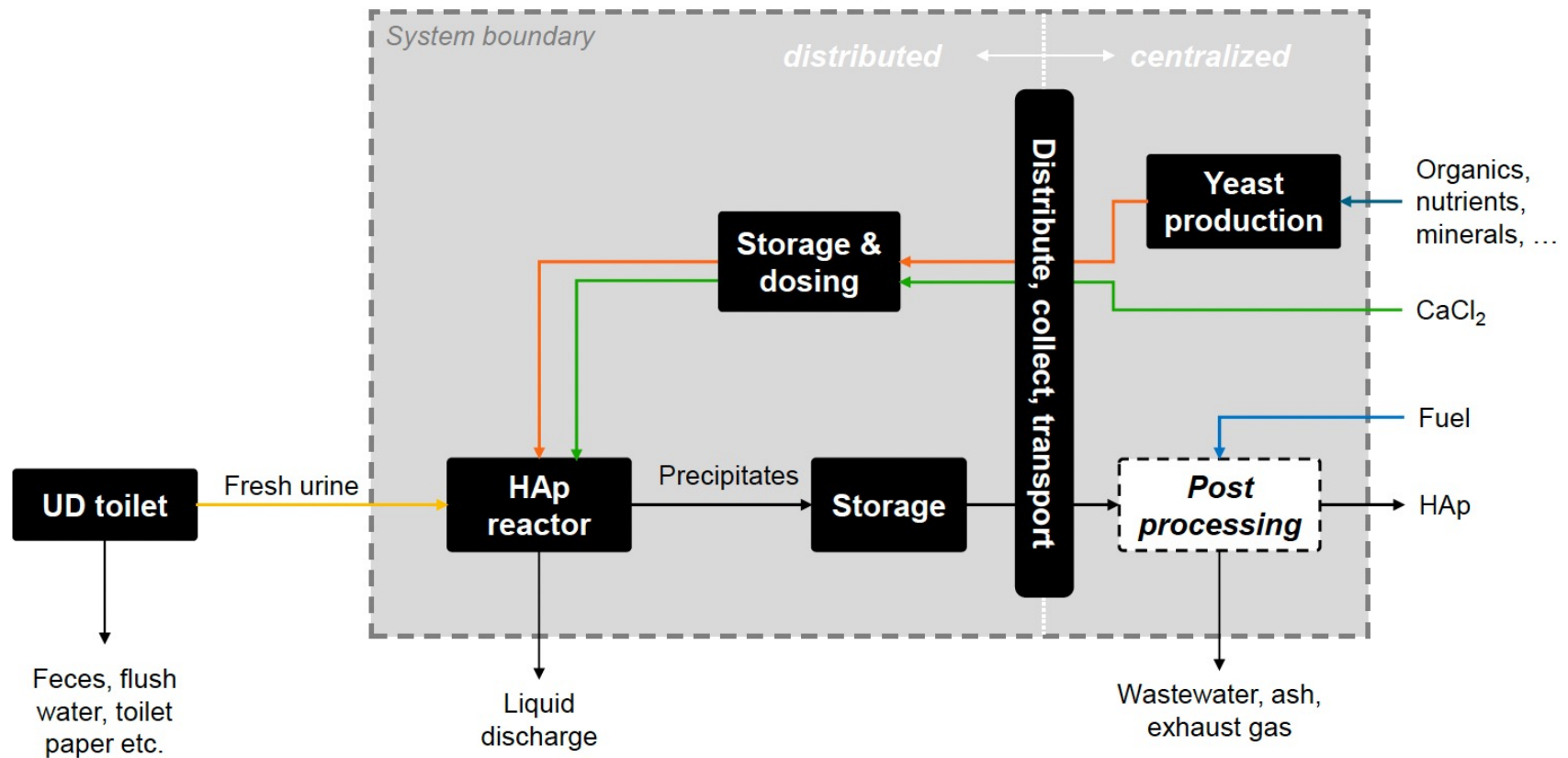
Technoeconomic assessment of HAp synthesis from Urine

- HAp production systems are deployed across a densely populated city, e.g., San Francisco
- Potential locations (📍) for deployment have consistently large fluxes of urine, e.g., schools, shopping malls
- Osteoeyeast are produced at a central location (★) and supplied to the distributed HAp reactors regularly.
- Precipitates will be collected regularly and further processed at the same central location (★).
- Toilets capable of urine diversion are assumed to be readily available at the deployed locations.

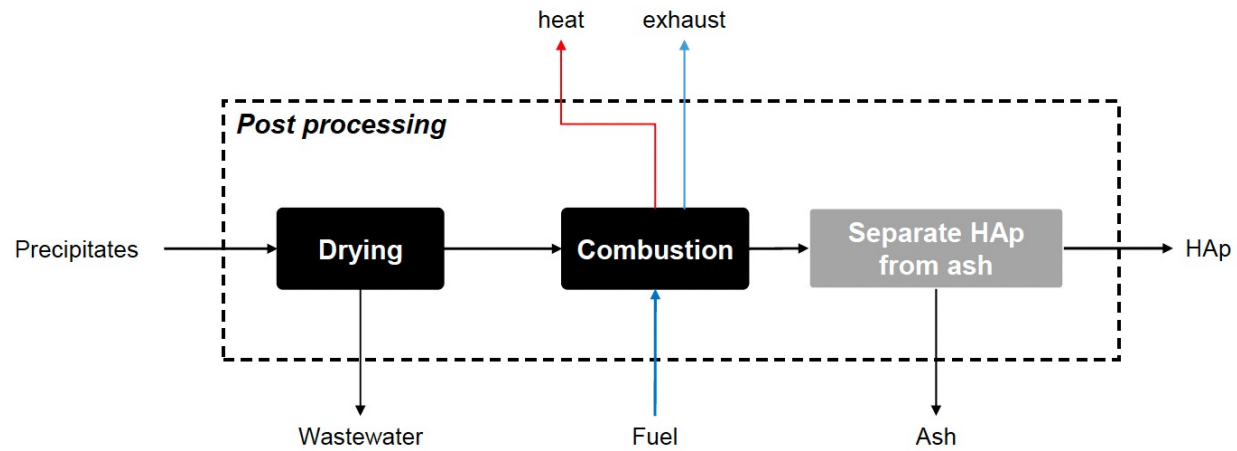


- San Francisco Unified School District

Technoeconomic assessment of HAp synthesis from Urine

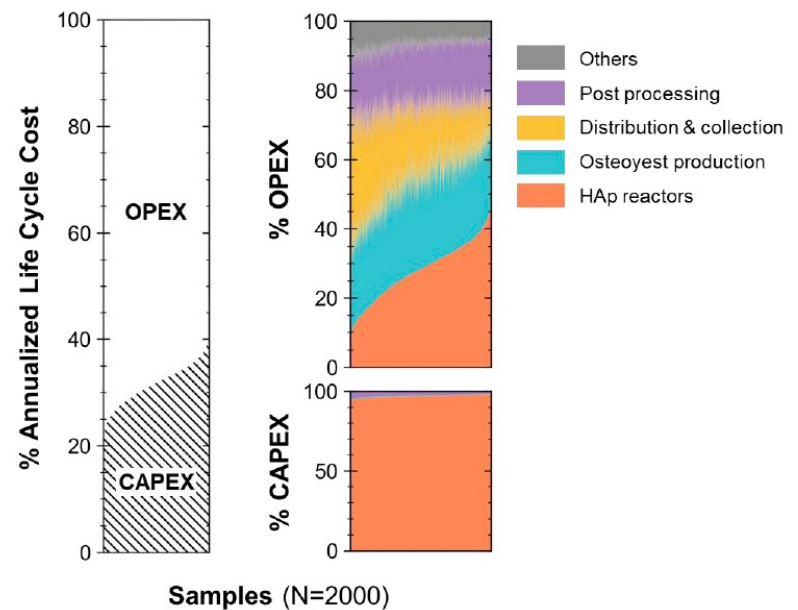
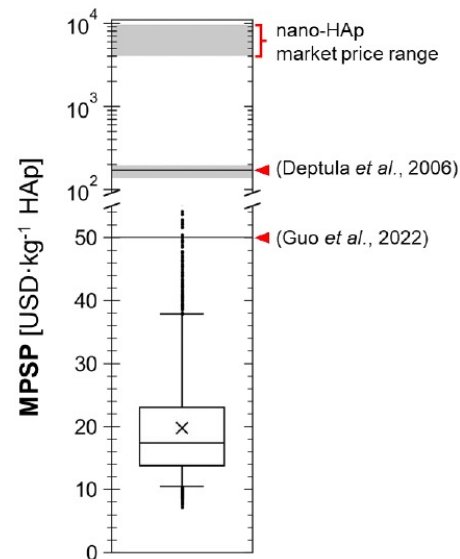


Technoeconomic assessment of HAp synthesis from Urine



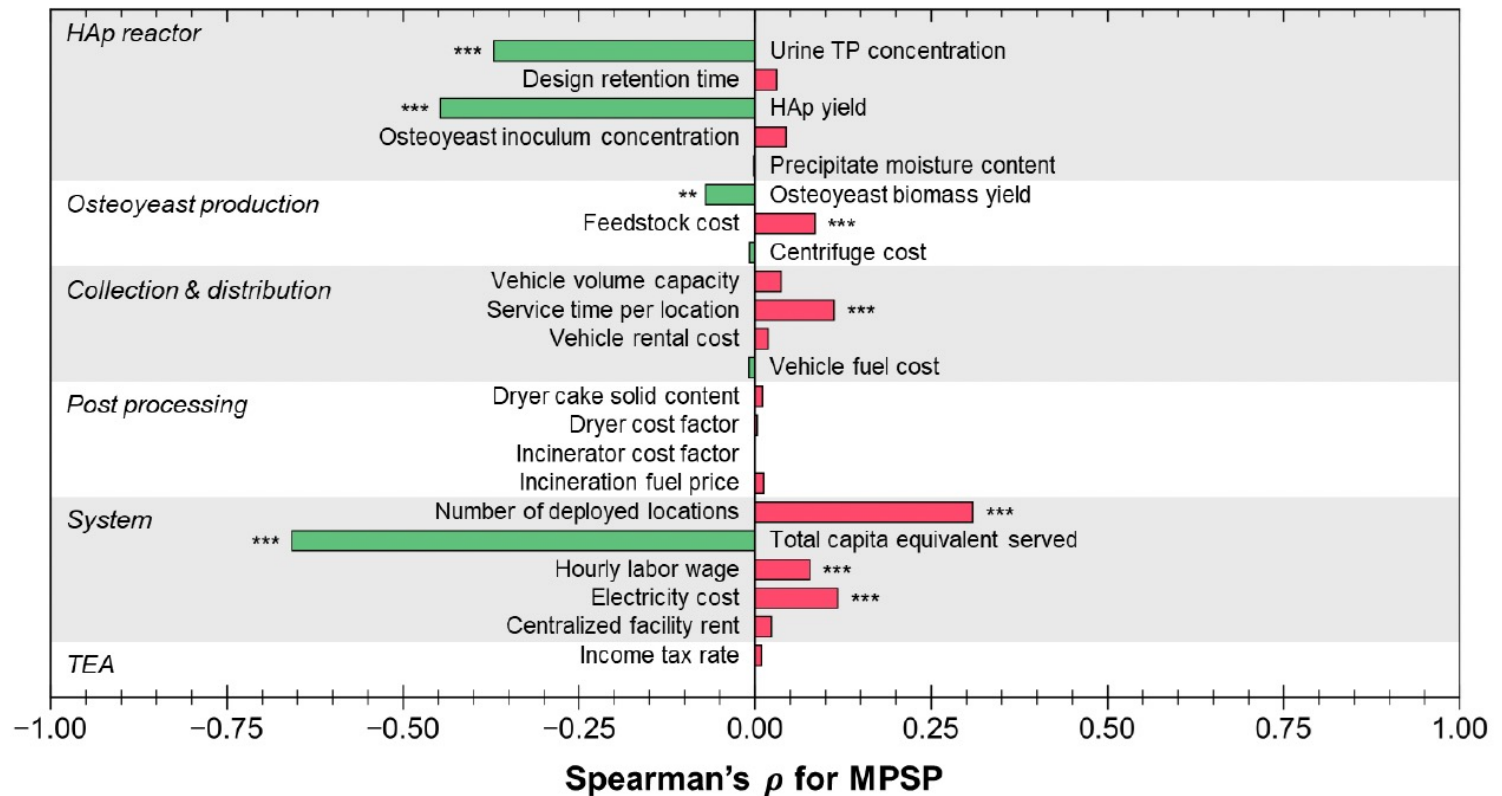
Technoeconomic assessment of HAp synthesis from Urine

- Other TEA assumptions
 - Project lifetime = 10 years
 - Discount rate = 0.05
- TEA indicators
 - Minimum product selling price to break even (**MPSP**, in USD/kg HAp)
 - Annualized life cycle cost
 - **CAPEX** (capital expenditure)
 - **OPEX** (operation expenditure)



Technoeconomic assessment of HAp synthesis from Urine

MPSP (Minimum Product Selling Price, in USD/kg HAp)



Summary

- **Introduced a new yellow carbon concept**
- **The osteoyeast platform designed based on osteoblast can produce Hap directly from urine with a high yield**
- **Osteoyeast-based HAp production makes the urine diversion process profitable**

Acknowledgements



Synthetic Biology Group

- Dr. Ian Blaby
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- Prof. Paul Jensen



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Please contact me if you are interested in a Post-Doc position